

**UTILITY
PATENT APPLICATION
TRANSMITTAL**

(Only for new nonprovisional applications under 37 CFR 1.53(b))

Attorney Docket No. CR9981 US DIV

First Named Inventor or Application Identifier

NAIR ET AL.

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Express Mailing Date OCTOBER 25, 2000

PRO

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09/655

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10/25/00

APPLICATION ELEMENTS

See MPEP chapter 600 concerning utility patent application contents.

ADDRESS TO: Assistant Commissioner for Patents
Box Patent Application
Washington, DC 20231

1. Fee (Authority to charge deposit account below.)
(Submit an original, and a duplicate for fee processing)

2. Specification [Total Pages 71]
(preferred arrangement set forth below)

- Descriptive title of the invention
- Cross References to Related Applications (if needed)
- Statement Regarding Fed sponsored R & D (if needed)
- Reference to Microfiche Appendix (if filed)
- Background of the Invention
- Brief Summary of the Invention
- Brief Description of the Drawings (if filed)
- Detailed Description
- Claim(s)
- Abstract of the Disclosure

6. Microfiche Computer Program (Appendix)

7. Nucleotide and/or Amino Acid Sequence Submission
(if applicable, all necessary)

- a. Computer Readable Copy
- b. Paper Copy (identical to computer copy)
(from parent case)
- c. Statement verifying identity of above copies

ACCOMPANYING APPLICATION PARTS

8. Power of Attorney

9. Information Disclosure Statement (IDS)/Cover Letter plus PTO-1449 Copies of IDS Citations

10. Preliminary Amendment

11. Return Receipt Postcard (MPEP 503)
(Should be specifically itemized)

12. Certified Copy of Priority Document(s)
(if foreign priority is claimed)

13. Other: Declaration under Budapest Treaty and ATCC receipt

3. Drawing(s) (35 USC 113) [Total Sheets 1]4. Oath or Declaration [Total Pages 2]

a. Newly executed (original or copy)

b. Copy from a prior application (37 CFR 1.63(d))
(for continuation/divisional with Box 14 completed)

- i. **DELETION OF INVENTORS**
- Signed Statement below at 15 deleting inventor(s) named in the prior application, see 37 CFR 1.63(d)(2) and 1.33(b).

5. Incorporation by Reference (useable if Box 4b is checked)
The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.

14. If a CONTINUING APPLICATION, check appropriate box and supply the requisite information:

Continuation Divisional Continuation-in-part (CIP) of prior Application No. 08/982,783

15. **DELETION OF INVENTOR(S) STATEMENT:** This application is being filed by less than all the inventors named in the prior application. In accordance with 37 CFR 1.63(d)(2) and 1.33(b), the Assistant Commissioner is requested to delete the name(s) of the following person or persons who are not inventors of the invention being claimed in this application:

16. Amend the specification by inserting before the first line the sentence:

-- This is a division of Application No. 08/982,783 filed December 3, 1997, which in turn is a continuation-in-part of U.S. Application No. 08/968,418, filed 12 November 1997, claiming priority to U.S. provisional Application No. 60/030,602, filed 13 November 1996. --

17. Cancel in this application original claims 17-20 of the prior application before calculating the filing leaving claims 1-16 and new claim 21 is added. (At least one original independent claim must be retained for filing purposes.)18. Priority of foreign Application No. _____ filed on _____ in _____

_____ is claimed under 35 U.S.C. 119.
(country)

CLAIMS	(1) FOR	(2) NUMBER FILED	(3) NUMBER EXTRA	(4) RATE	(5) CALCULATIONS
	TOTAL CLAIMS (37 CFR 1.16(c))	17 - 20 =	0	x \$ 18 =	0
	INDEPENDENT CLAIMS (37 CFR 1.16(b))	5 - 3 =	2	x \$ 80 =	\$ 160.00
	MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$ 270 =	0
				BASIC FEE (37 CFR 1.16(a))	+ \$ 710.00
				TOTAL =	\$ 870.00

19. The Commissioner is hereby authorized to credit overpayments or charge the following fees to Deposit Account No. 04-1928:

a. Fees required under 37 CFR 1.16.

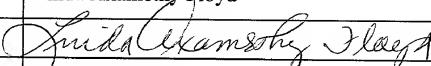
b. Fees required under 37 CFR 1.17.

20. Other:

21. CORRESPONDENCE ADDRESS

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22. SIGNATURE OF ATTORNEY OR AGENT REQUIRED

NAME	Linda Axamethy Floyd	REG. NO.: 33,692
SIGNATURE		
DATE	25 October 2000	

PATENT
EXPRESS MAIL LABEL NO.: EL073739163US

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In the Application of:

NAIR ET AL. CASE NO.: CR9981 US DIV

APPLICATION NO.: UNKNOWN GROUP ART UNIT: UNKNOWN

FILED: CONCURRENTLY HEREWITH EXAMINER: UNKNOWN

FOR: METHOD FOR THE PRODUCTION OF
GLYCEROL BY RECOMBINANT ORGANISMS

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
Washington, DC 20231

Sir:

Before examination of the above-referenced application,
please amend the application as follows:

IN THE SPECIFICATION:

On page 1, after the title, insert:

"This application is a division of 08/982,783 filed 3 December 1997, which in turn is a continuation-in-part of U.S. Application No. 08/968,418 filed 12 November 1997, claiming priority to U.S. Provisional Application No. 60/030,602, filed 13 November 1996."

At page 5, line 24, delete "12301 Parklawn Drive,
Rockville, MD 20852 U.S.A." and insert therefore --10801
University Blvd, Manassas, VA 20110-2209 U.S.A.--.

IN THE CLAIMS:

Add new Claim 21:

--21. A method for the production of glycerol from a recombinant organism comprising:

(i) transforming a suitable host cell with an expression cassette comprising either one or both of

(a) a gene encoding a glycerol-3-phosphate dehydrogenase enzyme selected from the group of enzymes consisting of sn-glycerol-3-phosphate:NAD⁺2-oxidoreductase and sn-glycerol-3-phosphate:NADP⁺2-oxidoreductase;

(b) a gene encoding a glycerol-3-phosphate phosphatase enzyme;

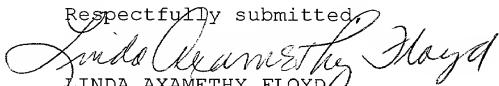
(ii) culturing the transformed host cell of (i) in the presence of at least one carbon source selected from the group consisting of monosaccharides, oligosaccharides, polysaccharides, and single-carbon substrates, whereby glycerol is produced; and

(iii) recovering the glycerol produced in (ii).-

REMARKS

Claims 1-16 and new Claim 21 are pending in this divisional application following entry of the above amendments.

Respectfully submitted


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Dated: 25 October 2000

TITLEMETHOD FOR THE PRODUCTION OF GLYCEROL
BY RECOMBINANT ORGANISMS
FIELD OF INVENTION

5 The present invention relates to the field of molecular biology and the use of recombinant organisms for the production of glycerol and compounds derived from the glycerol biosynthetic pathway. More specifically the invention describes the construction of a recombinant cell for the production of glycerol and derived compounds from a carbon substrate, the cell containing foreign 10 genes encoding proteins having glycerol-3-phosphate dehydrogenase (G3PDH) and glycerol-3-phosphatase (G3P phosphatase) activities where the endogenous genes encoding the glycerol-converting glycerol kinase and glycerol dehydrogenase activities have been deleted.

BACKGROUND

15 Glycerol is a compound in great demand by industry for use in cosmetics, liquid soaps, food, pharmaceuticals, lubricants, anti-freeze solutions, and in numerous other applications. The esters of glycerol are important in the fat and oil industry. Historically, glycerol has been isolated from animal fat and similar sources; however, the process is laborious and inefficient. Microbial 20 production of glycerol is preferred.

Not all organisms have a natural capacity to synthesize glycerol. However, the biological production of glycerol is known for some species of bacteria, algae, and yeast. The bacteria *Bacillus licheniformis* and *Lactobacillus lycopersica* synthesize glycerol. Glycerol production is found in the halotolerant 25 algae *Dunaliella sp.* and *Asteromonas gracilis* for protection against high external salt concentrations (Ben-Amotz et al., (1982) *Experientia* 38:49-52). Similarly, various osmotolerant yeast synthesize glycerol as a protective measure. Most strains of *Saccharomyces* produce some glycerol during 30 alcoholic fermentation and this production can be increased by the application of osmotic stress (Albertyn et al., (1994) *Mol. Cell. Biol.* 14, 4135-4144). Earlier this century glycerol was produced commercially with *Saccharomyces* cultures to which steering reagents were added such as sulfites or alkalis. Through the formation of an inactive complex, the steering agents block or inhibit the conversion of acetaldehyde to ethanol; thus, excess reducing equivalents 35 (NADH) are available to or "steered" towards dihydroxyacetone phosphate (DHAP) for reduction to produce glycerol. This method is limited by the partial inhibition of yeast growth that is due to the sulfites. This limitation can be partially overcome by the use of alkalis which create excess NADH equivalents by a different mechanism. In this practice, the alkalis initiated a Cannizarro

disproportionation to yield ethanol and acetic acid from two equivalents of acetaldehyde. Thus, although production of glycerol is possible from naturally occurring organisms, production is often subject to the need to control osmotic stress of the cultures and the production of sulfites. A method free from these 5 limitations is desirable. Production of glycerol from recombinant organisms containing foreign genes encoding key steps in the glycerol biosynthetic pathway is one possible route to such a method.

A number of the genes involved in the glycerol biosynthetic pathway have been isolated. For example, the gene encoding glycerol-3-phosphate 10 dehydrogenase (DAR1, GPD1) has been cloned and sequenced from *Saccharomyces diastaticus* (Wang et al., (1994), *J. Bact.* 176:7091-7095). The DAR1 gene was cloned into a shuttle vector and used to transform *E. coli* where expression produced active enzyme. Wang et al., *supra*, recognizes that DAR1 is regulated by the cellular osmotic environment but does not suggest how the 15 gene might be used to enhance glycerol production in a recombinant organism.

Other glycerol-3-phosphate dehydrogenase enzymes have been isolated. For example, sn-glycerol-3-phosphate dehydrogenase has been cloned and 20 sequenced from *S. cerevisiae* (Larason et al., (1993) *Mol. Microbiol.*, 10:1101). Albertyn et al., (1994) *Mol. Cell. Biol.*, 14:4135 teach the cloning of GPD1 encoding a glycerol-3-phosphate dehydrogenase from *S. cerevisiae*. Like Wang et al., both Albertyn et al. and Larason et al. recognize the osmo-sensitivity of 25 the regulation of this gene but do not suggest how the gene might be used in the production of glycerol in a recombinant organism.

As with G3PDH, glycerol-3-phosphatase has been isolated from 25 *Saccharomyces cerevisiae* and the protein identified as being encoded by the GPP1 and GPP2 genes (Norbeck et al., (1996) *J. Biol. Chem.*, 271:13875). Like the genes encoding G3PDH, it appears that GPP2 is osmotically-induced.

Although the genes encoding G3PDH and G3P phosphatase have been isolated, there is no teaching in the art that demonstrates glycerol production 30 from recombinant organisms with G3PDH/G3P phosphatase expressed together or separately. Further, there is no teaching to suggest that efficient glycerol production from any wild-type organism is possible using these two enzyme activities that does not require applying some stress (salt or an osmolyte) to the cell. In fact, the art suggests that G3PDH activities may not affect glycerol 35 production. For example, Eustace ((1987), *Can. J. Microbiol.*, 33:112-117)) teaches hybridized yeast strains that produced glycerol at greater levels than the parent strains. However, Eustace also demonstrates that G3PDH activity remained constant or slightly lower in the hybridized strains as opposed to the wild type.

Glycerol is an industrially useful material. However, other compounds may be derived from the glycerol biosynthetic pathway that also have commercial significance. For example, glycerol-producing organisms may be engineered to produce 1,3-propanediol (U.S. 5686276), a monomer having potential utility in the production of polyester fibers and the manufacture of polyurethanes and cyclic compounds. It is known for example that in some organisms, glycerol is converted to 3-hydroxypropionaldehyde and then to 1,3-propanediol through the actions of a dehydratase enzyme and an oxidoreductase enzyme, respectively. Bacterial strains able to produce 1,3-propanediol have been found, for example, in the groups *Citrobacter*, *Clostridium*, *Enterobacter*, *Ilyobacter*, *Klebsiella*, *Lactobacillus*, and *Pelobacter*. Glycerol dehydratase and diol dehydratase systems are described by Seyfried et al. (1996) *J. Bacteriol.* 178:5793-5796 and Tobimatsu et al. (1995) *J. Biol. Chem.* 270:7142-7148, respectively. Recombinant organisms, containing exogenous dehydratase enzyme, that are able to produce 1,3-propanediol have been described (U.S. 5686276). Although these organisms produce 1,3-propanediol, it is clear that they would benefit from a system that would minimize glycerol conversion.

There are a number of advantages in engineering a glycerol-producing organism for the production of 1,3-propanediol where conversion of glycerol is minimized. A microorganism capable of efficiently producing glycerol under physiological conditions is industrially desirable, especially when the glycerol itself will be used as a substrate *in vivo* as part of a more complex catabolic or biosynthetic pathway that could be perturbed by osmotic stress or the addition of steering agents (e.g., the production of 1,3-propanediol). Some attempts at creating glycerol kinase and glycerol dehydrogenase mutants have been made. For example, De Koning et al. (1990) *Appl. Microbiol Biotechnol.* 32:693-698 report the methanol-dependent production of dihydroxyacetone and glycerol by mutants of the methylotrophic yeast *Hansenula polymorpha* blocked in dihydroxyacetone kinase and glycerol kinase. Methanol and an additional substrate, required to replenish the xylose-5-phosphate co-substrate of the assimilation reaction, were used to produce glycerol; however, a dihydroxyacetone reductase (glycerol dehydrogenase) is also required. Similarly, Shaw and Cameron, Book of Abstracts, 211th ACS National Meeting, New Orleans, LA, March 24-28 (1996), BIOT-154 Publisher: American Chemical Society, Washington, D. C., investigate the deletion of *ldhA* (lactate dehydrogenase), *glpK* (glycerol kinase), and *tpiA* (triosephosphate isomerase) for the optimization of 1,3-propanediol production. They do not suggest the expression of cloned genes for G3PDH or G3P phosphatase for the

production of glycerol or 1,3-propanediol and they do not discuss the impact of glycerol dehydrogenase.

The problem to be solved, therefore, is the lack of a process to direct carbon flux towards glycerol production by the addition or enhancement of 5 certain enzyme activities, especially G3PDH and G3P phosphatase which respectively catalyze the conversion of dihydroxyacetone phosphate (DHAP) to glycerol-3-phosphate (G3P) and then to glycerol. The problem is complicated by the need to control the carbon flux away from glycerol by deletion or 10 decrease of certain enzyme activities, especially glycerol kinase and glycerol dehydrogenase which respectively catalyze the conversion of glycerol plus ATP to G3P and glycerol to dihydroxyacetone (or glyceraldehyde).

SUMMARY OF THE INVENTION

The present invention provides a method for the production of glycerol from a recombinant organism comprising: transforming a suitable host cell with 15 an expression cassette comprising either one or both of (a) a gene encoding a protein having glycerol-3-phosphate dehydrogenase activity and (b) a gene encoding a protein having glycerol-3-phosphate phosphatase activity, where the suitable host cell contains a disruption in either one or both of (a) a gene encoding an endogenous glycerol kinase and (b) a gene encoding an endogenous 20 glycerol dehydrogenase, wherein the disruption prevents the expression of active gene product; culturing the transformed host cell in the presence of at least one carbon source selected from the group consisting of monosaccharides, oligosaccharides, polysaccharides, and single-carbon substrates, whereby glycerol is produced; and recovering the glycerol produced.

The present invention further provides a process for the production of 25 1,3-propanediol from a recombinant organism comprising: transforming a suitable host cell with an expression cassette comprising either one or both of (a) a gene encoding a protein having glycerol-3-phosphate dehydrogenase activity and (b) a gene encoding a protein having glycerol-3-phosphate 30 phosphatase activity, the suitable host cell having at least one gene encoding a protein having a dehydratase activity and having a disruption in either one or both of (a) a gene encoding an endogenous glycerol kinase and (b) a gene encoding an endogenous glycerol dehydrogenase, wherein the disruption in the genes of (a) or (b) prevents the expression of active gene product; culturing the 35 transformed host cell in the presence of at least one carbon source selected from the group consisting of monosaccharides, oligosaccharides, polysaccharides, and single-carbon substrates whereby 1,3-propanediol is produced; and recovering the 1,3-propanediol produced.

Additionally, the invention provides for a process for the production of 1,3-propanediol from a recombinant organism where multiple copies of endogenous genes are introduced.

Further embodiments of the invention include host cells transformed with 5 heterologous genes for the glycerol pathway as well as host cells which contain endogenous genes for the glycerol pathway.

10 Additionally, the invention provides recombinant cells suitable for the production either glycerol or 1,3-propanediol, the host cells having genes expressing either one or both of a glycerol-3-phosphate dehydrogenase activity and a glycerol-3-phosphate phosphatase activity wherein the cell also has 15 disruptions in either one or both of a gene encoding an endogenous glycerol kinase and a gene encoding an endogenous glycerol dehydrogenase, wherein the disruption in the genes prevents the expression of active gene product.

BRIEF DESCRIPTION OF THE FIGURES, BIOLOGICAL
DEPOSITS AND SEQUENCE LISTING

15 Figure 1 illustrates the representative enzymatic pathways involving glycerol metabolism.

Applicants have made the following biological deposits under the terms of the Budapest Treaty on the International Recognition of the Deposit of 20 Micro-organisms for the Purposes of Patent Procedure:

Depositor Identification Reference	Int'l. Depository Designation	Date of Deposit
<i>Escherichia coli</i> pAH21/DH5 α (containing the GPP2 gene)	ATCC 98187	26 September 1996
<i>Escherichia coli</i> (pDAR1A/AA200) (containing the DAR1 gene)	ATCC 98248	6 November 1996
<i>FM5 Escherichia coli</i> RJF10m (containing a <i>glpK</i> disruption)	ATCC 98597	25 November 1997
<i>FM5 Escherichia coli</i> MSP33.6 (containing a <i>gldA</i> disruption)	ATCC 98598	25 November 1997

25 "ATCC" refers to the American Type Culture Collection international depository located at 12301 Parklawn Drive, Rockville, MD 20852 U.S.A. The designation is the accession number of the deposited material.

Applicants have provided 43 sequences in conformity with the Rules for the Standard Representation of Nucleotide and Amino Acid Sequences in Patent Applications (Annexes I and II to the Decision of the President of the EPO, published in Supplement No. 2 to OJ EPO, 12/1992) and with 37 C.F.R. 30 1.821-1.825 and Appendices A and B (Requirements for Application Disclosures Containing Nucleotides and/or Amino Acid Sequences).

DETAILED DESCRIPTION OF THE INVENTION

The present invention solves the problem stated above by providing a method for the biological production of glycerol from a fermentable carbon source in a recombinant organism. The method provides a rapid, inexpensive and environmentally-responsible source of glycerol useful in the cosmetics and pharmaceutical industries. The method uses a microorganism containing cloned homologous or heterologous genes encoding glycerol-3-phosphate dehydrogenase (G3PDH) and/or glycerol-3-phosphatase (G3P phosphatase). These genes are expressed in a recombinant host having disruptions in genes encoding endogenous glycerol kinase and/or glycerol dehydrogenase enzymes. The method is useful for the production of glycerol, as well as any end products for which glycerol is an intermediate. The recombinant microorganism is contacted with a carbon source and cultured and then glycerol or any end products derived therefrom are isolated from the conditioned media. The genes may be incorporated into the host microorganism separately or together for the production of glycerol.

Applicants' process has not previously been described for a recombinant organism and required the isolation of genes encoding the two enzymes and their subsequent expression in a host cell having disruptions in the endogenous kinase and dehydrogenase genes. It will be appreciated by those familiar with this art that Applicants' process may be generally applied to the production compounds where glycerol is a key intermediate, e.g., 1,3-propanediol.

As used herein the following terms may be used for interpretation of the claims and specification.

The terms "glycerol-3-phosphate dehydrogenase" and "G3PDH" refer to a polypeptide responsible for an enzyme activity that catalyzes the conversion of dihydroxyacetone phosphate (DHAP) to glycerol-3-phosphate (G3P). *In vivo* G3PDH may be NADH, NADPH; or FAD-dependent. The NADH-dependent enzyme (EC 1.1.1.8) is encoded, for example, by several genes including GPD1 (GenBank Z74071x2), or GPD2 (GenBank Z35169x1), or GPD3 (GenBank G984182), or DAR1 (GenBank Z74071x2). The NADPH-dependent enzyme (EC 1.1.1.94) is encoded by *gpsA* (GenBank U321643, (cds 197911-196892) G466746 and L45246). The FAD-dependent enzyme (EC 1.1.99.5) is encoded by GUT2 (GenBank Z47047x23), or *glpD* (GenBank G147838), or *glpABC* (GenBank M20938).

The terms "glycerol-3-phosphatase", "sn-glycerol-3-phosphatase", or "d,l-glycerol phosphatase", and "G3P phosphatase" refer to a polypeptide responsible for an enzyme activity that catalyzes the conversion of glycerol-3-phosphate and water to glycerol and inorganic phosphate. G3P phosphatase is

encoded, for example, by GPP1 (GenBank Z47047x125), or GPP2 (GenBank U18813x11).

5 The term "glycerol kinase" refers to a polypeptide responsible for an enzyme activity that catalyzes the conversion of glycerol and ATP to glycerol-3-phosphate and ADP. The high energy phosphate donor ATP may be replaced by physiological substitutes (e.g. phosphoenolpyruvate). Glycerol kinase is encoded, for example, by GUT1 (GenBank U11583x19) and *glpK* (GenBank L19201).

10 The term "glycerol dehydrogenase" refers to a polypeptide responsible for an enzyme activity that catalyzes the conversion of glycerol to dihydroxyacetone (E.C. 1.1.1.6) or glycerol to glyceraldehyde (E.C. 1.1.1.72). A polypeptide responsible for an enzyme activity that catalyzes the conversion of glycerol to dihydroxyacetone is also referred to as a "dihydroxyacetone 15 reductase". Glycerol dehydrogenase may be dependent upon NADH (E.C. 1.1.1.6), NADPH (E.C. 1.1.1.72), or other cofactors (e.g., E.C. 1.1.99.22). A NADH-dependent glycerol dehydrogenase is encoded, for example, by *gldA* (GenBank U00006).

20 The term "dehydratase enzyme" will refer to any enzyme that is capable of isomerizing or converting a glycerol molecule to the product 3-hydroxypropion-aldehyde. For the purposes of the present invention the dehydratase enzymes include a glycerol dehydratase (E.C. 4.2.1.30) and a diol dehydratase (E.C. 4.2.1.28) having preferred substrates of glycerol and 25 1,2-propanediol, respectively. In *Citrobacter freundii*, for example, glycerol dehydratase is encoded by three polypeptides whose gene sequences are represented by *dhaB*, *dhaC* and *dhaE* (GenBank U09771: base pairs 8556-10223, 10235-10819, and 10822-11250, respectively). In *Klebsiella oxytoca*, for example, diol dehydratase is encoded by three polypeptides whose gene sequences are represented by *pddA*, *pddB*, and *pddC* (GenBank D45071: base pairs 121-1785, 1796-2470, and 2485-3006, respectively).

30 The terms "GPD1", "DAR1", "OSG1", "D2830", and "YDL022W" will be used interchangeably and refer to a gene that encodes a cytosolic glycerol-3-phosphate dehydrogenase and is characterized by the base sequence given as SEQ ID NO:1.

35 The term "GPD2" refers to a gene that encodes a cytosolic glycerol-3-phosphate dehydrogenase and is characterized by the base sequence given in SEQ ID NO:2.

The terms "GUT2" and "YIL155C" are used interchangeably and refer to a gene that encodes a mitochondrial glycerol-3-phosphate dehydrogenase and is characterized by the base sequence given in SEQ ID NO:3.

The terms "GPP1", "RHR2" and "YIL053W" are used interchangeably and refer to a gene that encodes a cytosolic glycerol-3-phosphatase and is characterized by the base sequence given in SEQ ID NO:4.

5 The terms "GPP2", "HOR2" and "YER062C" are used interchangeably and refer to a gene that encodes a cytosolic glycerol-3-phosphatase and is characterized by the base sequence given as SEQ ID NO:5.

The term "GUT1" refers to a gene that encodes a cytosolic glycerol kinase and is characterized by the base sequence given as SEQ ID NO:6. The term "*glpK*" refers to another gene that encodes a glycerol kinase and is 10 characterized by the base sequence given in GeneBank L19201, base pairs 77347-78855.

The term "*gld4*" refers to a gene that encodes a glycerol dehydrogenase and is characterized by the base sequence given in GeneBank U00006, base pairs 3174-4316. The term "*dhaD*" refers to another gene that encodes a 15 glycerol dehydrogenase and is characterized by the base sequence given in GeneBank U09771, base pairs 2557-3654.

As used herein, the terms "function" and "enzyme function" refer to the catalytic activity of an enzyme in altering the energy required to perform a specific chemical reaction. Such an activity may apply to a reaction in 20 equilibrium where the production of both product and substrate may be accomplished under suitable conditions.

The terms "polypeptide" and "protein" are used interchangeably.

The terms "carbon substrate" and "carbon source" refer to a carbon source capable of being metabolized by host organisms of the present invention 25 and particularly mean carbon sources selected from the group consisting of monosaccharides, oligosaccharides, polysaccharides, and one-carbon substrates or mixtures thereof.

"Conversion" refers to the metabolic processes of an organism or cell that by means of a chemical reaction degrades or alters the complexity of a 30 chemical compound or substrate.

The terms "host cell" and "host organism" refer to a microorganism capable of receiving foreign or heterologous genes and additional copies of endogenous genes and expressing those genes to produce an active gene product.

35 The terms "production cell" and "production organism" refer to a cell engineered for the production of glycerol or compounds that may be derived from the glycerol biosynthetic pathway. The production cell will be recombinant and contain either one or both of a gene that encodes a protein having a glycerol-3-phosphate dehydrogenase activity and a gene encoding a

protein having a glycerol-3-phosphatase activity. In addition to the G3PDH and G3P phosphatase genes, the host cell will contain disruptions in one or both of a gene encoding an endogenous glycerol kinase and a gene encoding an endogenous glycerol dehydrogenase. Where the production cell is designed to 5 produce 1,3-propanediol, it will additionally contain a gene encoding a protein having a dehydratase activity.

The terms “foreign gene”, “foreign DNA”, “heterologous gene”, and “heterologous DNA” all refer to genetic material native to one organism that has been placed within a different host organism.

10 The term “endogenous” as used herein with reference to genes or polypeptides expressed by genes, refers to genes or polypeptides that are native to a production cell and are not derived from another organism. Thus an “endogenous glycerol kinase” and an “endogenous glycerol dehydrogenase” are terms referring to polypeptides encoded by genes native to the production cell.

15 The terms “recombinant organism” and “transformed host” refer to any organism transformed with heterologous or foreign genes. The recombinant organisms of the present invention express foreign genes encoding G3PDH and G3P phosphatase for the production of glycerol from suitable carbon substrates. Additionally, the terms “recombinant organism” and “transformed host” refer to 20 any organism transformed with endogenous (or homologous) genes so as to increase the copy number of the genes.

25 “Gene” refers to a nucleic acid fragment that expresses a specific protein, including regulatory sequences preceding (5' non-coding) and following (3' non-coding) the coding region. The terms “native” and “wild-type” gene refer to the gene as found in nature with its own regulatory sequences.

The terms “encoding” and “coding” refer to the process by which a 30 gene, through the mechanisms of transcription and translation, produces an amino acid sequence. The process of encoding a specific amino acid sequence is meant to include DNA sequences that may involve base changes that do not cause a change in the encoded amino acid, or which involve base changes which 35 may alter one or more amino acids, but do not affect the functional properties of the protein encoded by the DNA sequence. Therefore, the invention encompasses more than the specific exemplary sequences. Modifications to the sequence, such as deletions, insertions, or substitutions in the sequence which produce silent changes that do not substantially affect the functional properties of the resulting protein molecule are also contemplated. For example, alterations in the gene sequence which reflect the degeneracy of the genetic code, or which result in the production of a chemically equivalent amino acid at a given site, are contemplated; thus, a codon for the amino acid alanine, a

hydrophobic amino acid, may be substituted by a codon encoding another less hydrophobic residue, such as glycine, or a more hydrophobic residue, such as valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for glutamic acid, or one positively charged residue for another, such as lysine for arginine, can also be expected to produce a biologically equivalent product. Nucleotide changes which result in alteration of the N-terminal and C-terminal portions of the protein molecule would also not be expected to alter the activity of the protein. In some cases, it may in fact be desirable to make mutants of the sequence in order to study the effect of alteration on the biological activity of the protein. Each of the proposed modifications is well within the routine skill in the art, as is determination of retention of biological activity in the encoded products. Moreover, the skilled artisan recognizes that sequences encompassed by this invention are also defined by their ability to hybridize, under stringent conditions (0.1X SSC, 0.1% SDS, 65 °C), with the sequences exemplified herein.

The term "expression" refers to the transcription and translation to gene product from a gene coding for the sequence of the gene product.

The terms "plasmid", "vector", and "cassette" as used herein refer to an extra chromosomal element often carrying genes which are not part of the central metabolism of the cell and usually in the form of circular double-stranded DNA molecules. Such elements may be autonomously replicating sequences, genome integrating sequences, phage or nucleotide sequences, linear or circular, of a single- or double-stranded DNA or RNA, derived from any source, in which a number of nucleotide sequences have been joined or recombined into a unique construction which is capable of introducing a promoter fragment and DNA sequence for a selected gene product along with appropriate 3' untranslated sequence into a cell. "Transformation cassette" refers to a specific vector containing a foreign gene and having elements in addition to the foreign gene that facilitate transformation of a particular host cell. "Expression cassette" refers to a specific vector containing a foreign gene and having elements in addition to the foreign gene that allow for enhanced expression of that gene in a foreign host.

The terms "transformation" and "transfection" refer to the acquisition of new genes in a cell after the incorporation of nucleic acid. The acquired genes may be integrated into chromosomal DNA or introduced as extrachromosomal replicating sequences. The term "transformant" refers to the cell resulting from a transformation.

The term "genetically altered" refers to the process of changing hereditary material by transformation or mutation. The terms "disruption" and "gene interrupt" as applied to genes refer to a method of genetically altering an organism by adding to or deleting from a gene a significant portion of that gene such that the protein encoded by that gene is either not expressed or not expressed in active form.

Glycerol Biosynthetic Pathway

It is contemplated that glycerol may be produced in recombinant organisms by the manipulation of the glycerol biosynthetic pathway found in most microorganisms. Typically, a carbon substrate such as glucose is converted to glucose-6-phosphate via hexokinase in the presence of ATP. Glucose-phosphate isomerase catalyzes the conversion of glucose-6-phosphate to fructose-6-phosphate and then to fructose-1,6-diphosphate through the action of 6-phosphofructokinase. The diphosphate is then taken to dihydroxyacetone phosphate (DHAP) via aldolase. Finally NADH-dependent G3PDH converts DHAP to glycerol-3-phosphate which is then dephosphorylated to glycerol by G3P phosphatase. (Agarwal (1990), *Adv. Biochem. Engrg.* 41:114).

Genes encoding G3PDH, glycerol dehydrogenase, G3P phosphatase and glycerol kinase

The present invention provides genes suitable for the expression of G3PDH and G3P phosphatase activities in a host cell.

Genes encoding G3PDH are known. For example, GPD1 has been isolated from *Saccharomyces* and has the base sequence given by SEQ ID NO:1, encoding the amino acid sequence given in SEQ ID NO:7 (Wang et al., *supra*).

Similarly, G3PDH activity has also been isolated from *Saccharomyces* encoded by GPD2 having the base sequence given in SEQ ID NO:2 encoding the amino acid sequence given in SEQ ID NO:8 (Eriksson et al., (1995) *Mol. Microbiol.*, 17:95).

For the purposes of the present invention it is contemplated that any gene encoding a polypeptide responsible for G3PDH activity is suitable wherein that activity is capable of catalyzing the conversion of dihydroxyacetone phosphate (DHAP) to glycerol-3-phosphate (G3P). Further, it is contemplated that any gene encoding the amino acid sequence of G3PDH as given by SEQ ID NOS:7, 8, 9, 10, 11 and 12 corresponding to the genes GPD1, GPD2, GUT2, gpsA, glpD, and the α subunit of glpABC respectively, will be functional in the present invention wherein that amino acid sequence may encompass amino acid substitutions, deletions or additions that do not alter the function of the enzyme. The skilled person will appreciate that genes encoding G3PDH isolated from other sources will also be suitable for use in the present invention. For

example, genes isolated from prokaryotes include GenBank accessions M34393, M20938, L06231, U12567, L45246, L45323, L45324, L45325, U32164, U32689, and U39682. Genes isolated from fungi include GenBank accessions U30625, U30876 and X56162; genes isolated from insects include GenBank accessions X61223 and X14179; and genes isolated from mammalian sources include GenBank accessions U12424, M25558 and X78593.

5 Genes encoding G3P phosphatase are known. For example, GPP2 has been isolated from *Saccharomyces cerevisiae* and has the base sequence given by SEQ ID NO:5, which encodes the amino acid sequence given in SEQ ID NO:13 (Norbeck et al., (1996), *J. Biol. Chem.*, 271:13875).

10 For the purposes of the present invention, any gene encoding a G3P phosphatase activity is suitable for use in the method wherein that activity is capable of catalyzing the conversion of glycerol-3-phosphate and water to glycerol and inorganic phosphate. Further, any gene encoding the amino acid 15 sequence of G3P phosphatase as given by SEQ ID NOS:13 and 14 corresponding to the genes GPP2 and GPP1 respectively, will be functional in the present invention including any amino acid sequence that encompasses amino acid substitutions, deletions or additions that do not alter the function of the G3P phosphatase enzyme. The skilled person will appreciate that genes encoding 20 G3P phosphatase isolated from other sources will also be suitable for use in the present invention. For example, the dephosphorylation of glycerol-3-phosphate to yield glycerol may be achieved with one or more of the following general or specific phosphatases: alkaline phosphatase (EC 3.1.3.1) [GenBank M19159, M29663, U02550 or M33965]; acid phosphatase (EC 3.1.3.2) [GenBank U51210, U19789, U28658 or L20566]; glycerol-3-phosphatase (EC 3.1.3.-) 25 [GenBank Z38060 or U18813x11]; glucose-1-phosphatase (EC 3.1.3.10) [GenBank M33807]; glucose-6-phosphatase (EC 3.1.3.9) [GenBank U00445]; fructose-1,6-bisphosphatase (EC 3.1.3.11) [GenBank X12545 or J03207] or phosphotidyl glycerol phosphate phosphatase (EC 3.1.3.27) [GenBank M23546 30 and M23628].

Genes encoding glycerol kinase are known. For example, GUT1 encoding the glycerol kinase from *Saccharomyces* has been isolated and sequenced (Pavlik et al. (1993), *Curr. Genet.*, 24:21) and the base sequence is given by SEQ ID NO:6, which encodes the amino acid sequence given in 35 SEQ ID NO:15. Alternatively, *glpK* encodes a glycerol kinase from *E. coli* and is characterized by the base sequence given in GeneBank L19201, base pairs 77347-78855.

Genes encoding glycerol dehydrogenase are known. For example, *gldA* encodes a glycerol dehydrogenase from *E. coli* and is characterized by the base

sequence given in GeneBank U00006, base pairs 3174-4316. Alternatively, *dhAD* refers to another gene that encodes a glycerol dehydrogenase from *Citrobacter freundii* and is characterized by the base sequence given in GeneBank U09771, base pairs 2557-3654.

5 Host cells

Suitable host cells for the recombinant production of glycerol by the expression of G3PDH and G3P phosphatase may be either prokaryotic or eukaryotic and will be limited only by their ability to express active enzymes. Preferred host cells will be those bacteria, yeasts, and filamentous fungi.

10 typically useful for the production of glycerol such as *Citrobacter*, *Enterobacter*, *Clostridium*, *Klebsiella*, *Aerobacter*, *Lactobacillus*, *Aspergillus*, *Saccharomyces*, *Schizosaccharomyces*, *Zygosaccharomyces*, *Pichia*, *Kluyveromyces*, *Candida*, *Hansenula*, *Debaryomyces*, *Mucor*, *Torulopsis*, *Methylobacter*, *Escherichia*, *Salmonella*, *Bacillus*, *Streptomyces* and *Pseudomonas*. Preferred in the present

15 invention are *E. coli* and *Saccharomyces*.

Where glycerol is a key intermediate in the production of 1,3-propanediol the host cell will either have an endogenous gene encoding a protein having a dehydratase activity or will acquire such a gene through transformation. Host cells particularly suited for production of 1,3-propanediol are *Citrobacter*, *Enterobacter*, *Clostridium*, *Klebsiella*, *Aerobacter*, *Lactobacillus*, and *Salmonella*, which have endogenous genes encoding dehydratase enzymes. Additionally, host cells that lack such an endogenous gene include *E. coli*.

Vectors And Expression Cassettes

The present invention provides a variety of vectors and transformation and expression cassettes suitable for the cloning, transformation and expression of G3PDH and G3P phosphatase into a suitable host cell. Suitable vectors will be those which are compatible with the bacterium employed. Suitable vectors can be derived, for example, from a bacteria, a virus (such as bacteriophage T7 or a M-13 derived phage), a cosmid, a yeast or a plant. Protocols for obtaining and using such vectors are known to those in the art (Sambrook et al., Molecular Cloning: A Laboratory Manual - volumes 1, 2, 3 (Cold Spring Harbor Laboratory: Cold Spring Harbor, NY, 1989)).

Typically, the vector or cassette contains sequences directing transcription and translation of the appropriate gene, a selectable marker, and sequences allowing autonomous replication or chromosomal integration. Suitable vectors comprise a region 5' of the gene which harbors transcriptional initiation controls and a region 3' of the DNA fragment which controls transcriptional termination. It is most preferred when both control regions are derived from genes homologous to the transformed host cell. Such control

regions need not be derived from the genes native to the specific species chosen as a production host.

Initiation control regions, or promoters, which are useful to drive expression of the G3PDH and G3P phosphatase genes in the desired host cell are numerous and familiar to those skilled in the art. Virtually any promoter capable of driving these genes is suitable for the present invention including but not limited to CYC1, HIS3, GAL1, GAL10, ADH1, PGK, PHO5, GAPDH, ADC1, TRP1, URA3, LEU2, ENO, and TPI (useful for expression in *Saccharomyces*); AOX1 (useful for expression in *Pichia*); and lac, trp, λ P_L, λ P_R, T7, tac, and trc, (useful for expression in *E. coli*).

Termination control regions may also be derived from various genes native to the preferred hosts. Optionally, a termination site may be unnecessary; however, it is most preferred if included.

For effective expression of the instant enzymes, DNA encoding the enzymes are linked operably through initiation codons to selected expression control regions such that expression results in the formation of the appropriate messenger RNA.

Transformation Of Suitable Hosts And Expression Of G3PDH And G3P Phosphatase For The Production Of Glycerol

Once suitable cassettes are constructed they are used to transform appropriate host cells. Introduction of the cassette containing the genes encoding G3PDH and/or G3P phosphatase into the host cell may be accomplished by known procedures such as by transformation, e.g., using calcium-permeabilized cells, electroporation, or by transfection using a recombinant phage virus (Sambrook et al., *supra*).

In the present invention AH21 and DAR1 cassettes were used to transform the *E. coli* DH5 α and FM5 as fully described in the GENERAL METHODS and EXAMPLES.

Random And Site Specific Mutagenesis For Disrupting Enzyme Activities:

Enzyme pathways by which organisms metabolize glycerol are known in the art, Figure 1. Glycerol is converted to glycerol-3-phosphate (G3P) by an ATP-dependent glycerol kinase; the G3P may then be oxidized to DHAP by G3PDH. In a second pathway, glycerol is oxidized to dihydroxyacetone (DHA) by a glycerol dehydrogenase; the DHA may then be converted to DHAP by an ATP-dependent DHA kinase. In a third pathway, glycerol is oxidized to glyceraldehyde by a glycerol dehydrogenase; the glyceraldehyde may be phosphorylated to glyceraldehyde-3-phosphate by an ATP-dependent kinase. DHAP and glyceraldehyde-3-phosphate, interconverted by the action of triosephosphate isomerase, may be further metabolized via central metabolism

pathways. These pathways, by introducing by-products, are deleterious to glycerol production.

One aspect of the present invention is the ability to provide a production organism for the production of glycerol where the glycerol-converting activities of glycerol kinase and glycerol dehydrogenase have been deleted. Methods of creating deletion mutants are common and well known in the art. For example, wild type cells may be exposed to a variety of agents such as radiation or chemical mutagens and then screened for the desired phenotype. When creating mutations through radiation either ultraviolet (UV) or ionizing radiation may be used. Suitable short wave UV wavelengths for genetic mutations will fall within the range of 200 nm to 300 nm where 254 nm is preferred. UV radiation in this wavelength principally causes changes within nucleic acid sequence from guanine and cytosine to adenine and thymidine. Since all cells have DNA repair mechanisms that would repair most UV induced mutations, agents such as caffeine and other inhibitors may be added to interrupt the repair process and maximize the number of effective mutations. Long wave UV mutations using light in the 300 nm to 400 nm range are also possible but are generally not as effective as the short wave UV light unless used in conjunction with various activators such as psoralen dyes that interact with the DNA.

Mutagenesis with chemical agents is also effective for generating mutants and commonly used substances include chemicals that affect nonreplicating DNA such as HNO_2 and NH_2OH , as well as agents that affect replicating DNA such as acridine dyes, notable for causing frameshift mutations. Specific methods for creating mutants using radiation or chemical agents are well documented in the art. See for example Thomas D. Brock in *Biotechnology: A Textbook of Industrial Microbiology*, Second Edition (1989) Sinauer Associates, Inc., Sunderland, MA., or Deshpande, Mukund V., *Appl. Biochem. Biotechnol.*, 36, 227, (1992), herein incorporated by reference.

After mutagenesis has occurred, mutants having the desired phenotype may be selected by a variety of methods. Random screening is most common where the mutagenized cells are selected for the ability to produce the desired product or intermediate. Alternatively, selective isolation of mutants can be performed by growing a mutagenized population on selective media where only resistant colonies can develop. Methods of mutant selection are highly developed and well known in the art of industrial microbiology. See Brock, *Supra.*, DeMancilha et al., *Food Chem.*, 14, 313, (1984).

Biological mutagenic agents which target genes randomly are well known in the art. See for example De Bruijn and Rossbach in *Methods for General and Molecular Bacteriology* (1994) American Society for Microbiology,

Washington, D.C. Alternatively, provided that gene sequence is known, chromosomal gene disruption with specific deletion or replacement is achieved by homologous recombination with an appropriate plasmid. See for example Hamilton et al. (1989) *J. Bacteriol.* 171:4617-4622, Balbas et al. (1993) *Gene* 136: 211-213, Gueldener et al. (1996) *Nucleic Acids Res.* 24: 2519-2524, and Smith et al. (1996) *Methods Mol. Cell. Biol.* 5: 270-277.

5 It is contemplated that any of the above cited methods may be used for the deletion or inactivation of glycerol kinase and glycerol dehydrogenase activities in the preferred production organism.

10 Media and Carbon Substrates

Fermentation media in the present invention must contain suitable carbon substrates. Suitable substrates may include but are not limited to mono-saccharides such as glucose and fructose, oligosaccharides such as lactose or sucrose, polysaccharides such as starch or cellulose or mixtures thereof and 15 unpurified mixtures from renewable feedstocks such as cheese whey permeate, cornsteep liquor, sugar beet molasses, and barley malt. Additionally, the carbon substrate may also be one-carbon substrates such as carbon dioxide, or methanol for which metabolic conversion into key biochemical intermediates has been demonstrated.

20 Glycerol production from single carbon sources (e.g., methanol, formaldehyde or formate) has been reported in methylotrophic yeasts (Yamada et al. (1989), *Agric. Biol. Chem.*, 53(2):541-543) and in bacteria (Hunter et al. (1985), *Biochemistry*, 24:4148-4155). These organisms can assimilate single carbon compounds, ranging in oxidation state from methane to formate, and 25 produce glycerol. The pathway of carbon assimilation can be through ribulose monophosphate, through serine, or through xylulose-monophosphate (Gottschalk, Bacterial Metabolism, Second Edition, Springer-Verlag: New York (1986)). The ribulose monophosphate pathway involves the condensation of formate with ribulose-5-phosphate to form a 6 carbon sugar that becomes 30 fructose and eventually the three carbon product, glyceraldehyde-3-phosphate. Likewise, the serine pathway assimilates the one-carbon compound into the glycolytic pathway via methylenetetrahydrofolate.

In addition to one and two carbon substrates, methylotrophic organisms are also known to utilize a number of other carbon-containing compounds such 35 as methylamine, glucosamine and a variety of amino acids for metabolic activity. For example, methylotrophic yeast are known to utilize the carbon from methylamine to form trehalose or glycerol (Bellion et al. (1993), *Microb. Growth C1 Compd.*, [Int. Symp.], 7th, 415-32. Editor(s): Murrell, J. Collin; Kelly, Don P. Publisher: Intercept, Andover, UK). Similarly, various species

of *Candida* will metabolize alanine or oleic acid (Sulter et al. (1990), *Arch. Microbiol.*, 153(5):485-9). Hence, the source of carbon utilized in the present invention may encompass a wide variety of carbon-containing substrates and will only be limited by the choice of organism.

5 Although all of the above mentioned carbon substrates and mixtures thereof are suitable in the present invention, preferred carbon substrates are monosaccharides, oligosaccharides, polysaccharides, single-carbon substrates or mixtures thereof. More preferred are sugars such as glucose, fructose, sucrose, maltose, lactose and single carbon substrates such as methanol and carbon dioxide. Most preferred as a carbon substrate is glucose.

10 In addition to an appropriate carbon source, fermentation media must contain suitable minerals, salts, cofactors, buffers and other components, known to those skilled in the art, suitable for the growth of the cultures and promotion of the enzymatic pathway necessary for glycerol production.

15 Culture Conditions

Typically cells are grown at 30 °C in appropriate media. Preferred growth media are common commercially prepared media such as Luria Bertani (LB) broth, Sabouraud Dextrose (SD) broth, or Yeast medium (YM) broth. Other defined or synthetic growth media may also be used and the appropriate medium for growth of the particular microorganism will be known by one skilled in the art of microbiology or fermentation science. The use of agents known to modulate catabolite repression directly or indirectly, e.g., cyclic adenosine 3':5'-monophosphate, may also be incorporated into the reaction media. Similarly, the use of agents known to modulate enzymatic activities (e.g., sulfites, bisulfites, and alkalis) that lead to enhancement of glycerol production may be used in conjunction with or as an alternative to genetic manipulations.

Suitable pH ranges for the fermentation are between pH 5.0 to pH 9.0 where the range of pH 6.0 to pH 8.0 is preferred for the initial condition.

30 Reactions may be performed under aerobic or anaerobic conditions where anaerobic or microaerobic conditions are preferred.

Identification of G3PDH, glycerol dehydrogenase, G3P phosphatase, and glycerol kinase activities

35 The levels of expression of the proteins G3PDH, G3P phosphatase, glycerol dehydrogenase, and glycerol kinase are measured by enzyme assays. Generally, G3PDH activity and glycerol dehydrogenase activity assays rely on the spectral properties of the cosubstrate, NADH, in the DHAP conversion to G-3-P and the DHA conversion to glycerol, respectively. NADH has intrinsic UV/vis absorption and its consumption can be monitored spectrophotometrically

at 340 nm. G3P phosphatase activity can be measured by any method of measuring the inorganic phosphate liberated in the reaction. The most commonly used detection method uses the visible spectroscopic determination of a blue-colored phosphomolybdate ammonium complex. Glycerol kinase activity

5 can be measured by the detection of G3P from glycerol and ATP, for example, by NMR. Assays can be directed toward more specific characteristics of individual enzymes if necessary, for example, by the use of alternate cofactors.

Identification and recovery of glycerol and other products (e.g. 1,3-propanediol)

10 Glycerol and other products (e.g. 1,3-propanediol) may be identified and quantified by high performance liquid chromatography (HPLC) and gas chromatography/mass spectroscopy (GC/MS) analyses on the cell-free extracts. Preferred is a HPLC method where the fermentation media are analyzed on an analytical ion exchange column using a mobile phase of 0.01N sulfuric acid in an isocratic fashion.

15 Methods for the recovery of glycerol from fermentation media are known in the art. For example, glycerol can be obtained from cell media by subjecting the reaction mixture to the following sequence of steps: filtration; water removal; organic solvent extraction; and fractional distillation (U.S. Patent No. 2,986,495).

20 Description Of The Preferred Embodiments

Production of Glycerol

25 The present invention describes a method for the production of glycerol from a suitable carbon source utilizing a recombinant organism. Particularly suitable in the invention is a bacterial host cell, transformed with an expression cassette carrying either or both of a gene that encodes a protein having a glycerol-3-phosphate dehydrogenase activity and a gene encoding a protein having a glycerol-3-phosphatase activity. In addition to the G3PDH and G3P phosphatase genes, the host cell will contain disruptions in either or both of genes encoding endogenous glycerol kinase and glycerol dehydrogenase. 30 The combined effect of the foreign G3PDH and G3P phosphatase genes (providing a pathway from the carbon source to glycerol) with the gene disruptions (blocking the conversion of glycerol) results in an organism that is capable of efficient and reliable glycerol production.

35 Although the optimal organism for glycerol production contains the above mentioned gene disruptions, glycerol production is possible with a host cell containing either one or both of the foreign G3PDH and G3P phosphatase genes in the absence of such disruptions. For example, the recombinant *E. coli* strain AA200 carrying the DAR1 gene (Example 1) was capable of producing between 0.38 g/L and 0.48 g/L of glycerol depending on fermentation

parameters. Similarly, the *E. coli* DH5 α , carrying and expressible GPP2 gene (Example 2), was capable of 0.2 g/L of glycerol production. Where both genes are present, (Example 3 and 4), glycerol production attained about 40 g/L. Where both genes are present in conjunction with an elimination of the

5 endogenous glycerol kinase activity, a reduction in the conversion of glycerol may be seen (Example 8). Furthermore, the presence of glycerol dehydrogenase activity is linked to the conversion of glycerol under glucose-limited conditions; thus, it is anticipated that the elimination of glycerol dehydrogenase activity will result in the reduction of glycerol conversion (Example 8).

10 **Production of 1,3-propanediol**

The present invention may also be adapted for the production of 1,3-propanediol by utilizing recombinant organisms expressing the foreign G3PDH and/or G3P phosphatase genes and containing disruptions in the endogenous glycerol kinase and/or glycerol dehydrogenase activities.

15 Additionally, the invention provides for the process for the production of 1,3-propanediol from a recombinant organism where multiple copies of endogenous genes are introduced. In addition to these genetic alterations, the production cell will require the presence of a gene encoding an active dehydratase enzyme. The dehydratase enzyme activity may either be a glycerol

20 dehydratase or a diol dehydratase. The dehydratase enzyme activity may result from either the expression of an endogenous gene or from the expression of a foreign gene transfected into the host organism. Isolation and expression of genes encoding suitable dehydratase enzymes are well known in the art and are taught by applicants in PCT/US96/06705, filed 5 November 1996 and

25 U.S. 5686276 and U.S. 5633362, hereby incorporated by reference. It will be appreciated that, as glycerol is a key intermediate in the production of 1,3-propanediol, where the host cell contains a dehydratase activity in conjunction with expressed foreign G3PDH and/or G3P phosphatase genes and in the absence of the glycerol-converting glycerol kinase or glycerol

30 dehydrogenase activities, the cell will be particularly suited for the production of 1,3-propanediol.

The present invention is further defined in the following Examples. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From the 35 above discussion and these Examples, one skilled in the art can ascertain the essential characteristics of this invention, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions.

EXAMPLES

GENERAL METHODS

Procedures for phosphorylations, ligations, and transformations are well known in the art. Techniques suitable for use in the following examples may be found in Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor Laboratory Press (1989).

Materials and methods suitable for the maintenance and growth of bacterial cultures are well known in the art. Techniques suitable for use in the following examples may be found in Manual of Methods for General

10 Bacteriology (Phillipp Gerhardt, R. G. E. Murray, Ralph N. Costilow, Eugene W. Nester, Willis A. Wood, Noel R. Krieg and G. Briggs Phillips, eds), American Society for Microbiology, Washington, DC. (1994) or in Biotechnology: A Textbook of Industrial Microbiology (Thomas D. Brock, Second Edition (1989) Sinauer Associates, Inc., Sunderland, MA). All reagents 15 and materials used for the growth and maintenance of bacterial cells were obtained from Aldrich Chemicals (Milwaukee, WI), DIFCO Laboratories (Detroit, MI), GIBCO/BRL (Gaithersburg, MD), or Sigma Chemical Company (St. Louis, MO) unless otherwise specified.

The meaning of abbreviations is as follows: "h" means hour(s), "min" means minute(s), "sec" means second(s), "d" means day(s), "mL" means milliliters, "L" means liters.

Cell strains

The following *Escherichia coli* strains were used for transformation and expression of G3PDH and G3P phosphatase. Strains were obtained from the 25 *E. coli* Genetic Stock Center, ATCC, or Life Technologies (Gaithersburg, MD).

AA200 (*garB10 fhuA22 ompF627 fadL701 relA1 pit-10 spot1 tpi-1 phoM510 mcrB1*) (Anderson et al., (1970), *J. Gen. Microbiol.*, 62:329).

30 BB20 (*tonA22 ΔphoA8 fadL701 relA1 glpR2 glpD3 pit-10 gpsA20 spot1 T2R*) (Cronan et al., *J. Bact.*, 118:598).

DH5α (*deoR endA1 gyrA96 hsdR17 recA1 relA1 supE44 thi-1 Δ(lacZYA- argFV169) phi80lacZΔM15 F-*) (Woodcock et al., (1989), *Nucl. Acids Res.*,

35 17:3469).

FM5 Escherichia coli (ATCC 53911)

Identification of Glycerol

The conversion of glucose to glycerol was monitored by HPLC and/or GC. Analyses were performed using standard techniques and materials available to one of skill in the art of chromatography. One suitable method utilized a 5 Waters Maxima 820 HPLC system using UV (210 nm) and RI detection. Samples were injected onto a Shodex SH-1011 column (8 mm x 300 mm; Waters, Milford, MA) equipped with a Shodex SH-1011P precolumn (6 mm x 50 mm), temperature-controlled at 50 °C, using 0.01 N H₂SO₄ as mobile phase at a flow rate of 0.69 mL/min. When quantitative analysis was desired, samples 10 were prepared with a known amount of trimethylacetic acid as an external standard. Typically, the retention times of 1,3-propanediol (RI detection), glycerol (RI detection) and glucose (RI detection) were 21.39 min, 17.03 min and 12.66 min, respectively.

Glycerol was also analyzed by GC/MS. Gas chromatography with mass 15 spectrometry detection for separation and quantitation of glycerol was performed using a DB-WAX column (30 m, 0.32 mm I.D., 0.25 um film thickness, J & W Scientific, Folsom, CA) at the following conditions: injector: split, 1:15; sample volume: 1 uL; temperature profile: 150 °C intitial temperature with 30 sec hold, 40 °C/min to 180 °C, 20 °C/min to 240 °C, hold for 2.5 min. 20 Detection: EI Mass Spectrometry (Hewlett Packard 5971, San Fernando, CA), quantitative SIM using ions 61 m/z and 64 m/z as target ions for glycerol and glycerol-d8, and ion 43 m/z as qualifier ion for glycerol. Glycerol-d8 was used as an internal standard.

Assay for glycerol-3-phosphatase, G3P phosphatase

25 The assay for enzyme activity was performed by incubating the extract with an organic phosphate substrate in a bis-Tris or MES and magnesium buffer, pH 6.5. The substrate used was either l- α -glycerol phosphate, or d,l- α -glycerol phosphate. The final concentrations of the reagents in the assay are: buffer (20 mM bis-Tris or 50 mM MES); MgCl₂ (10 mM); and substrate (20 mM). If 30 the total protein in the sample was low and no visible precipitation occurs with an acid quench, the sample was conveniently assayed in the cuvette. This method involved incubating an enzyme sample in a cuvette that contained 20 mM substrate (50 μ L, 200 mM), 50 mM MES, 10 mM MgCl₂, pH 6.5 buffer. The final phosphatase assay volume was 0.5 mL. The enzyme- 35 containing sample was added to the reaction mixture; the contents of the cuvette were mixed and then the cuvette was placed in a circulating water bath at T = 37 °C for 5 to 120 min, the length of time depending on whether the phosphatase activity in the enzyme sample ranged from 2 to 0.02 U/mL. The enzymatic reaction was quenched by the addition of the acid molybdate reagent

(0.4 mL). After the Fiske SubbaRow reagent (0.1 mL) and distilled water (1.5 mL) were added, the solution was mixed and allowed to develop. After 10 min, to allow full color development, the absorbance of the samples was read at 660 nm using a Cary 219 UV/Vis spectrophotometer. The amount of 5 inorganic phosphate released was compared to a standard curve that was prepared by using a stock inorganic phosphate solution (0.65 mM) and preparing 6 standards with final inorganic phosphate concentrations ranging from 0.026 to 0.130 μ mol/mL.

Spectrophotometric Assay for Glycerol 3-Phosphate Dehydrogenase (G3PDH)

10 Activity

The following procedure was used as modified below from a method published by Bell et al. (1975), *J. Biol. Chem.*, 250:7153-8. This method involved incubating an enzyme sample in a cuvette that contained 0.2 mM NADH; 2.0 mM dihydroxyacetone phosphate (DHAP), and enzyme in 0.1 M 15 Tris/HCl, pH 7.5 buffer with 5 mM DTT, in a total volume of 1.0 mL at 30 °C. The spectrophotometer was set to monitor absorbance changes at the fixed wavelength of 340 nm. The instrument was blanked on a cuvette containing buffer only. After the enzyme was added to the cuvette, an absorbance reading was taken. The first substrate, NADH (50 μ L 4 mM NADH; absorbance should 20 increase approx 1.25 AU), was added to determine the background rate. The rate should be followed for at least 3 min. The second substrate, DHAP (50 μ L 40 mM DHAP), was then added and the absorbance change over time was monitored for at least 3 min to determine to determine the gross rate. G3PDH activity was defined by subtracting the background rate from the gross rate.

25 ^{13}C -NMR Assay for Glycerol Kinase Activity

An appropriate amount of enzyme, typically a cell-free crude extract, was added to a reaction mixture containing 40 mM ATP, 20 mM MgSO_4 , 21 mM uniformly ^{13}C labelled glycerol (99%, Cambridge Isotope Laboratories), and 0.1 M Tris-HCl, pH 9 for 75 min at 25 °C. The conversion of glycerol to 30 glycerol 3-phosphate was detected by ^{13}C -NMR (125 MHz): glycerol (63.11 ppm, d, $J = 41$ Hz and 72.66 ppm, t, $J = 41$ Hz); glycerol 3-phosphate (62.93 ppm, d, $J = 41$ Hz; 65.31 ppm, br d, $J = 43$ Hz; and 72.66 ppm, dt, $J = 6, 41$ Hz).

NADH-linked Glycerol Dehydrogenase Assay

35 NADH -linked glycerol dehydrogenase activity in *E. coli* strains (*gldA*) was determined after protein separation by non-denaturing polyacrylamide gel electrophoresis. The conversion of glycerol plus NAD^+ to dihydroxyacetone plus NADH was coupled with the conversion of 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide (MTT) to a deeply colored formazan, using

Electrophoresis was performed in duplicate by standard procedures using native gels (8-16% TG, 1.5 mm, 15 lane gels from Novex, San Diego, CA).

5 Residual glycerol was removed from the gels by washing 3x with 50 mM Tris or potassium carbonate buffer, pH 9 for 10 min. The duplicate gels were developed, with and without glycerol (approx. 0.16 M final concentration), in 15 mL of assay solution containing 50 mM Tris or potassium carbonate, pH 9, 60 mg ammonium sulfate, 75 mg NAD⁺, 1.5 mg MTT, and 0.5 mg PMS.

10 The presence or absence of NADH-linked glycerol dehydrogenase activity in *E. coli* strains (*gldA*) was also determined, following polyacrylamide gel electrophoresis, by reaction with polyclonal antibodies raised to purified *K. pneumoniae* glycerol dehydrogenase (*dhaD*).

PLASMID CONSTRUCTION AND STRAIN CONSTRUCTION

15 **Cloning and expression of glycerol 3-phosphatase for increase of glycerol production in *E. coli* DH5 α and FMS**

The *Saccharomyces cerevisiae* chromosome V lambda clone 6592 (Gene Bank, accession # U18813x11) was obtained from ATCC. The glycerol 3-phosphate phosphatase (GPP2) gene was cloned by cloning from the lambda 20 clone as target DNA using synthetic primers (SEQ ID NO:16 with SEQ ID NO:17) incorporating an BamHI-RBS-XbaI site at the 5' end and a SmaI site at the 3' end. The product was subcloned into pCR-Script (Stratagene, Madison, WI) at the SrfI site to generate the plasmids pAH15 containing GPP2. The plasmid pAH15 contains the GPP2 gene in the inactive orientation for 25 expression from the lac promoter in pCR-Script SK+. The BamHI-SmaI fragment from pAH15 containing the GPP2 gene was inserted into pBlueScriptII SK+ to generate plasmid pAH19. The pAH19 contains the GPP2 gene in the correct orientation for expression from the lac promoter. The XbaI-PstI fragment from pAH19 containing the GPP2 gene was inserted into pPHOX2 to 30 create plasmid pAH21. The pAH21/ DH5 α is the expression plasmid.

Plasmids for the over-expression of DAR1 in *E. coli*

DAR1 was isolated by PCR cloning from genomic *S. cerevisiae* DNA using synthetic primers (SEQ ID NO:18 with SEQ ID NO:19). Successful PCR cloning places an NcoI site at the 5' end of DAR1 where the ATG within NcoI 35 is the DAR1 initiator methionine. At the 3' end of DAR1 a BamHI site is introduced following the translation terminator. The PCR fragments were digested with NcoI + BamHI and cloned into the same sites within the expression plasmid pTrc99A (Pharmacia, Piscataway, NJ) to give pDAR1A.

In order to create a better ribosome binding site at the 5' end of DAR1, an SpeI-RBS-NcoI linker obtained by annealing synthetic primers (SEQ ID NO:20 with SEQ ID NO:21) was inserted into the NcoI site of pDAR1A to create pAH40. Plasmid pAH40 contains the new RBS and DAR1 gene in the correct orientation for expression from the trc promoter of pTrc99A (Pharmacia, Piscataway, NJ). The NcoI-BamHI fragment from pDAR1A and a second set of SpeI-RBS-NcoI linker obtained by annealing synthetic primers (SEQ ID NO:22 with SEQ ID NO:23) was inserted into the SpeI-BamHI site of pBC-SK+ (Stratagene, Madison, WI) to create plasmid pAH42. The plasmid pAH42 contains a chloramphenicol resistant gene.

Construction of expression cassettes for DAR1 and GPP2

Expression cassettes for DAR1 and GPP2 were assembled from the individual DAR1 and GPP2 subclones described above using standard molecular biology methods. The BamHI-PstI fragment from pAH19 containing the ribosomal binding site (RBS) and GPP2 gene was inserted into pAH40 to create pAH43. The BamHI-PstI fragment from pAH19 containing the RBS and GPP2 gene was inserted into pAH42 to create pAH45.

The ribosome binding site at the 5' end of GPP2 was modified as follows. A BamHI-RBS-SpeI linker, obtained by annealing synthetic primers GATCCAGGAAACAGA (SEQ ID NO:24) with CTAGTCTGTTTCCTG (SEQ ID NO:25) to the XbaI-PstI fragment from pAH19 containing the GPP2 gene, was inserted into the BamHI-PstI site of pAH40 to create pAH48. Plasmid pAH48 contains the DAR1 gene, the modified RBS, and the GPP2 gene in the correct orientation for expression from the trc promoter of pTrc99A (Pharmacia, Piscataway, NJ).

Transformation of *E. coli*

All the plasmids described here were transformed into *E. coli* DH5 α or FM5 using standard molecular biology techniques. The transformants were verified by its DNA RFLP pattern.

EXAMPLE 1
PRODUCTION OF GLYCEROL FROM *E. COLI*
TRANSFORMED WITH G3PDH GENE

Media

Synthetic media was used for anaerobic or aerobic production of glycerol using *E. coli* cells transformed with pDAR1A. The media contained per liter 6.0 g Na₂HPO₄, 3.0 g KH₂PO₄, 1.0 g NH₄Cl, 0.5 g NaCl, 1 mL 20% MgSO₄·7H₂O, 8.0 g glucose, 40 mg casamino acids, 0.5 ml 1% thiamine hydrochloride, 100 mg ampicillin.

Growth Conditions

Strain AA200 harboring pDAR1A or the pTrc99A vector was grown in aerobic conditions in 50 mL of media shaking at 250 rpm in 250 mL flasks at 37 °C. At A_{600} 0.2-0.3 isopropylthio- β -D-galactoside was added to a final concentration of 1 mM and incubation continued for 48 h. For anaerobic growth samples of induced cells were used to fill Falcon #2054 tubes which were capped and gently mixed by rotation at 37 °C for 48 h. Glycerol production was determined by HPLC analysis of the culture supernatants. Strain pDAR1A/AA200 produced 0.38 g/L glycerol after 48 h under anaerobic conditions, and 0.48 g/L under aerobic conditions.

EXAMPLE 2

PRODUCTION OF GLYCEROL FROM *E. COLI* TRANSFORMED WITH G3P PHOSPHATASE GENE (GPP2)

Media

Synthetic phoA media was used in shake flasks to demonstrate the increase of glycerol by GPP2 expression in *E. coli*. The phoA medium contained per liter: Amisoy, 12 g; ammonium sulfate, 0.62 g; MOPS, 10.5 g; Na-citrate, 1.2 g; NaOH (1 M), 10 mL; 1 M MgSO₄, 12 mL; 100X trace elements, 12 mL; 50% glucose, 10 mL; 1% thiamine, 10 mL; 100 mg/mL L-proline, 10 mL; 2.5 mM FeCl₃, 5 mL; mixed phosphates buffer, 2 mL (5 mL 0.2 M NaH₂PO₄ + 9 mL 0.2 M K₂HPO₄), and pH to 7.0. The 100X traces elements for phoA medium /L contained: ZnSO₄·7 H₂O, 0.58 g; MnSO₄·H₂O, 0.34 g; CuSO₄·5 H₂O, 0.49 g; CoCl₂·6 H₂O, 0.47 g; H₃BO₃, 0.12 g; NaMoO₄·2 H₂O, 0.48 g.

Shake Flasks Experiments

The strains pAH21/DH5 α (containing GPP2 gene) and pPHOX2/DH5 α (control) were grown in 45 mL of media (phoA media, 50 μ g/mL carbenicillin, and 1 μ g/mL vitamin B₁₂) in a 250 mL shake flask at 37 °C. The cultures were grown under aerobic condition (250 rpm shaking) for 24 h. Glycerol production was determined by HPLC analysis of the culture supernatant. pAH21/DH5 α produced 0.2 g/L glycerol after 24 h.

EXAMPLE 3

PRODUCTION OF GLYCEROL FROM D-GLUCOSE USING RECOMBINANT *E. COLI* CONTAINING BOTH GPP2 AND DAR1

Growth for demonstration of increased glycerol production by *E. coli* DH5 α -containing pAH43 proceeds aerobically at 37 °C in shake-flask cultures (erlenmeyer flasks, liquid volume 1/5th of total volume).

Cultures in minimal media/1% glucose shake-flasks are started by inoculation from overnight LB/1% glucose culture with antibiotic selection.

Minimal media are: filter-sterilized defined media, final pH 6.8 (HCl), contained per liter: 12.6 g (NH₄)₂SO₄, 13.7 g K₂HPO₄, 0.2 g yeast extract (Difco), 1 g NaHCO₃, 5 mg vitamin B₁₂, 5 mL Modified Balch's Trace-Element Solution (the composition of which can be found in Methods for General and Molecular Bacteriology (P. Gerhardt et al., eds, p. 158, American Society for Microbiology, Washington, DC (1994)). The shake-flasks are incubated at 37 °C with vigorous shaking for overnight, after which they are sampled for GC analysis of the supernatant. The pAH43/DH5 α showed glycerol production of 3.8 g/L after 24 h.

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EXAMPLE 4

PRODUCTION OF GLYCEROL FROM D-GLUCOSE USING RECOMBINANT *E. COLI* CONTAINING BOTH GPP2 AND DAR1

Example 4 illustrates the production of glucose from the recombinant *E. coli* DH5 α /pAH48, containing both the GPP2 and DAR1 genes.

15

The strain DH5 α /pAH48 was constructed as described above in the GENERAL METHODS.

Pre-Culture

DH5 α /pAH48 were pre-cultured for seeding into a fermentation run. Components and protocols for the pre-culture are listed below.

20

Pre-Culture Media

	KH ₂ PO ₄	30.0 g/L
	Citric acid	2.0 g/L
	MgSO ₄ ·7H ₂ O	2.0 g/L
	98% H ₂ SO ₄	2.0 mL/L
25	Ferric ammonium citrate	0.3 g/L
	CaCl ₂ ·2H ₂ O	0.2 g/L
	Yeast extract	5.0 g/L
	Trace metals	5.0 mL/L
	Glucose	10.0 g/L
30	Carbenicillin	100.0 mg/L

The above media components were mixed together and the pH adjusted to 6.8 with NH₄OH. The media was then filter sterilized.

Trace metals were used according to the following recipe:

35	Citric acid, monohydrate	4.0 g/L
	MgSO ₄ ·7H ₂ O	3.0 g/L
	MnSO ₄ ·H ₂ O	0.5 g/L
	NaCl	1.0 g/L
	FeSO ₄ ·7H ₂ O	0.1 g/L
	CoCl ₂ ·6H ₂ O	0.1 g/L

	CaCl ₂	0.1 g/L
	ZnSO ₄ ·7H ₂ O	0.1 g/L
	CuSO ₄ ·5 H ₂ O	10 mg/L
	AlK(SO ₄) ₂ ·12H ₂ O	10 mg/L
5	H ₃ BO ₃	10 mg/L
	Na ₂ MoO ₄ ·2H ₂ O	10 mg/L
	NiSO ₄ ·6H ₂ O	10 mg/L
	Na ₂ SeO ₃	10 mg/L
	Na ₂ WO ₄ ·2H ₂ O	10 mg/L
10	Cultures were started from seed culture inoculated from 50 µL frozen stock (15% glycerol as cryoprotectant) to 600 mL medium in a 2-L Erlenmeyer flask. Cultures were grown at 30 °C in a shaker at 250 rpm for approximately 12 h and then used to seed the fermenter.	

Fermentation growth

15 Vessel

15-L stirred tank fermenter

Medium

	KH ₂ PO ₄	6.8 g/L
	Citric acid	2.0 g/L
20	MgSO ₄ ·7H ₂ O	2.0 g/L
	98% H ₂ SO ₄	2.0 mL/L
	Ferric ammonium citrate	0.3 g/L
	CaCl ₂ ·2H ₂ O	0.2 g/L
	Mazu DF204 antifoam	1.0 mL/L

25 The above components were sterilized together in the fermenter vessel. The pH was raised to 6.7 with NH₄OH. Yeast extract (5 g/L) and trace metals solution (5 mL/L) were added aseptically from filter sterilized stock solutions. Glucose was added from 60% feed to give final concentration of 10 g/L. Carbenicillin was added at 100 mg/L. Volume after inoculation was 6 L.

30 Environmental Conditions For Fermentation

The temperature was controlled at 36 °C and the air flow rate was controlled at 6 standard liters per minute. Back pressure was controlled at 0.5 bar. The agitator was set at 350 rpm. Aqueous ammonia was used to control pH at 6.7. The glucose feed (60% glucose monohydrate) rate was controlled to maintain excess glucose.

Results

The results of the fermentation run are given in Table 1.

Table 1

EFT (hr)	OD550 (AU)	[Glucose] (g/L)	[Glycerol] (g/L)	Total Glucose Fed (g)	Total Glycerol Produced (g)
0	0.8	9.3		25	
6	4.7	4.0	2.0	49	14
8	5.4	0	3.6	71	25
10	6.7	0.0	4.7	116	33
12	7.4	2.1	7.0	157	49
14.2	10.4	0.3	10.0	230	70
16.2	18.1	9.7	15.5	259	106
18.2	12.4	14.5		305	
20.2	11.8	17.4	17.7	353	119
22.2	11.0	12.6		382	
24.2	10.8	6.5	26.6	404	178
26.2	10.9	6.8		442	
28.2	10.4	10.3	31.5	463	216
30.2	10.2	13.1	30.4	493	213
32.2	10.1	8.1	28.2	512	196
34.2	10.2	3.5	33.4	530	223
36.2	10.1	5.8		548	
38.2	9.8	5.1	36.1	512	233

EXAMPLE 5

**ENGINEERING OF GLYCEROL KINASE MUTANTS OF *E. COLI* FM5
FOR PRODUCTION OF GLYCEROL FROM GLUCOSE**

5 **Construction of integration plasmid for glycerol kinase gene replacement in
E. coli FM5**

E. coli FM5 genomic DNA was prepared using the Puregene DNA Isolation Kit (Genta Systems, Minneapolis, MN). A 1.0 kb DNA fragment containing partial *glpF* and glycerol kinase (*glpK*) genes was amplified by PCR 10 (Mullis and Faloona, *Methods Enzymol.*, 155:335-350, 1987) from FM5 genomic DNA using primers SEQ ID NO:26 and SEQ ID NO:27. A 1.1 kb DNA fragment containing partial *glpK* and *glpX* genes was amplified by PCR from FM5 genomic DNA using primers SEQ ID NO:28 and SEQ ID NO:29. A *Mun*I site was incorporated into primer SEQ ID NO:28. The 5' end of primer 15 SEQ ID NO:28 was the reverse complement of primer SEQ ID NO:27 to enable subsequent overlap extension PCR. The gene splicing by overlap extension technique (Horton et al., *BioTechniques*, 8:528-535, 1990) was used to generate a 2.1 kb fragment by PCR using the above two PCR fragments as templates and

primers SEQ ID NO:26 and SEQ ID NO:29. This fragment represented a deletion of 0.8 kb from the central region of the 1.5 kb *glpK* gene. Overall, this fragment had 1.0 kb and 1.1 kb flanking regions on either side of the *Mun*I cloning site (within the partial *glpK*) to allow for chromosomal gene replacement 5 by homologous recombination.

The above 2.1 kb PCR fragment was blunt-ended (using mung bean nuclease) and cloned into the pCR-Blunt vector using the Zero Blunt PCR Cloning Kit (Invitrogen, San Diego, CA) to yield the 5.6 kb plasmid pRN100 containing kanamycin and Zeocin resistance genes. The 1.2 kb *Hinc*II fragment 10 from pLoxCat1 (unpublished results), containing a chloramphenicol-resistance gene flanked by bacteriophage P1 *loxP* sites (Snaith et al., *Gene*, 166:173-174, 1995), was used to interrupt the *glpK* fragment in plasmid pRN100 by ligating it to *Mun*I-digested (and blunt-ended) plasmid pRN100 to yield the 6.9 kb plasmid pRN101-1. A 376 bp fragment containing the R6K origin was amplified by 15 PCR from the vector pGP704 (Miller and Mekalanos, *J. Bacteriol.*, 170:2575-2583, 1988) using primers SEQ ID NO:30 and SEQ ID NO:31, blunt-ended, and ligated to the 5.3 kb *Asp*718-*Aat*II fragment (which was blunt-ended) from pRN101-1 to yield the 5.7 kb plasmid pRN102-1 containing 20 kanamycin and chloramphenicol resistance genes. Substitution of the ColE1 origin region in pRN101-1 with the R6K origin to generate pRN102-1 also involved deletion of most of the Zeocin resistance gene. The host for pRN102-1 replication was *E. coli* SY327 (Miller and Mekalanos, *J. Bacteriol.*, 170:2575-2583, 1988) which contains the *pir* gene necessary for the function of the R6K origin.

25 Engineering Of Glycerol Kinase Mutant RJF10m With Chloramphenicol Resistance Gene Interrupt

E. coli FM5 was electrotransformed with the non-replicative integration plasmid pRN102-1 and transformants that were chloramphenicol-resistant (12.5 μ g/mL) and kanamycin-sensitive (30 μ g/mL) were further screened for 30 glycerol non-utilization on M9 minimal medium containing 1 mM glycerol. An *Eco*RI digest of genomic DNA from one such mutant, RJF10m, when probed with the intact *glpK* gene via Southern analysis (Southern, *J. Mol. Biol.*, 98:503-517, 1975) indicated that it was a double-crossover integrant (*glpK* gene replacement) since the two expected 7.9 kb and 2.0 kb bands were observed, 35 owing to the presence of an additional *Eco*RI site within the chloramphenicol resistance gene. The wild-type control yielded the single expected 9.4 kb band. A ^{13}C NMR analysis of mutant RJF10m confirmed that it was incapable of converting ^{13}C -labeled glycerol and ATP to glycerol-3-phosphate. This *glpK* mutant was further analyzed by genomic PCR using primer combinations SEQ

ID NO:32 and SEQ ID NO:33, SEQ ID NO:34 and SEQ ID NO:35, and SEQ ID NO:32 and SEQ ID NO:35 which yielded the expected 2.3 kb, 2.4 kb, and 4.0 kb PCR fragments respectively. The wild-type control yielded the expected 3.5 kb band with primers SEQ ID NO:32 and SEQ ID NO:35. The *glpK* mutant

5 RJF10m was electrotransformed with plasmid pAH48 to allow glycerol production from glucose. The *glpK* mutant *E. coli* RJF10m has been deposited with ATCC under the terms of the Budapest Treaty on 24 November 1997.

Engineering Of Glycerol Kinase Mutant RJF10 With Chloramphenicol Resistance Gene Interrupt Removed

10 After overnight growth on YENB medium (0.75 % yeast extract, 0.8% nutrient broth) at 37 °C, *E. coli* RJF10m in a water suspension was electrotransformed with plasmid pJW168 (unpublished results), which contained the bacteriophage P1 Cre recombinase gene under the control of the IPTG-inducible *lacUV5* promoter, a temperature-sensitive pSC101 replicon, and an ampicillin resistance gene. Upon outgrowth in SOC medium at 30 °C, transformants were selected at 30 °C (permissive temperature for pJW168 replication) on LB agar medium supplemented with carbenicillin (50 µg/mL) and IPTG (1 mM). Two serial overnight transfers of pooled colonies were carried out at 30 °C on fresh LB agar medium supplemented with carbenicillin and 15 IPTG in order to allow excision of the chromosomal chloramphenicol resistance gene via recombination at the *loxP* sites mediated by the Cre recombinase (Hoess and Abremski, *J. Mol. Biol.*, 181:351-362, 1985). Resultant colonies were replica-plated on to LB agar medium supplemented with carbenicillin and IPTG and LB agar supplemented with chloramphenicol (12.5 µg/mL) to identify 20 colonies that were carbenicillin-resistant and chloramphenicol-sensitive indicating marker gene removal. An overnight 30 °C culture of one such colony was used to inoculate 10 mL of LB medium. Upon growth at 30 °C to OD (600 nm) of 0.6, the culture was incubated at 37 °C overnight. Several dilutions were plated on prewarmed LB agar medium and the plates incubated overnight 25 at 42 °C (the non-permissive temperature for pJW168 replication). Resultant colonies were replica-plated on to LB agar medium and LB agar medium supplemented with carbenicillin (75 µg/mL) to identify colonies that were carbenicillin-sensitive indicating loss of plasmid pJW168. One such *glpK* mutant, RJF10, was further analyzed by genomic PCR using primers SEQ ID 30 NO:32 and SEQ ID NO:35 and yielded the expected 3.0 kb band confirming marker gene excision. Glycerol non-utilization by mutant RJF10 was confirmed by lack of growth on M9 minimal medium containing 1 mM glycerol. The *glpK* mutant RJF10 was electrotransformed with plasmid pAH48 to allow glycerol 35 production from glucose.

EXAMPLE 6

CONSTRUCTION OF *E. COLI* STRAIN WITH *GLDA* GENE KNOCKOUT

The *glcA* gene was isolated from *E. coli* by PCR (K. B. Mullis and F. A. Falloona (1987) *Meth. Enzymol.* 155:335-350) using primers SEQ ID NO:36 and SEQ ID NO:37, which incorporate terminal Sph1 and Xba1 sites, respectively, and cloned (T. Maniatis 1982 *Molecular Cloning. A Laboratory Manual*. Cold Spring Harbor, Cold Spring Harbor, NY) between the Sph1 and Xba1 sites in pUC18, to generate pKP8. pKP8 was cut at the unique Sal1 and Nco1 sites within the *glcA* gene, the ends flushed with Klenow and religated, resulting in a 109 bp deletion in the middle of *glcA* and regeneration of a unique Sal1 site, to generate pKP9. A 1.4 kb DNA fragment containing the gene conferring kanamycin resistance (kan), and including about 400 bps of DNA upstream of the translational start codon and about 100 bps of DNA downstream of the translational stop codon, was isolated from pET-28a(+) (Novagen, 10 Madison, Wis) by PCR using primers SEQ ID NO:38 and SEQ ID NO:39, which incorporate terminal Sal1 sites, and subcloned into the unique Sal1 site of pKP9, to generate pKP13. A 2.1 kb DNA fragment beginning 204 bps downstream of the *glcA* translational start codon and ending 178 bps upstream of the *glcA* translational stop codon, and containing the kan insertion, was isolated from pKP13 by PCR using primers SEQ ID NO:40 and SEQ ID NO:41, which incorporate terminal Sph1 and Xba1 sites, respectively, was subcloned between the Sph1 and Xba1 sites in pMAK705 (Genencor International, Palo Alto, Calif.), to generate pMP33. *E. coli* FM5 was transformed with pMP33 and selected on 20 ug/mL kan at 30 °C, which is the permissive temperature for pMAK705 replication. One colony was expanded overnight at 30 °C in liquid media supplemented with 20 ug/mL kan. Approximately 32,000 cells were plated on 20 ug/mL kan and incubated for 16 hrs at 44 °C, which is the restrictive temperature for pMAK705 replication. Transformants growing at 44 °C have plasmid integrated into the chromosome, occurring at a frequency of approximately 0.0001. PCR and Southern blot (E.M. Southern 1975 *J. Mol. Biol.* 98:503-517) analyses were used to determine the nature of the chromosomal integration events in the transformants. Western blot analysis (H. Towbin, et al. (1979) *Proc. Natl. Acad. Sci.* 76:4350) was used to determine whether glycerol dehydrogenase protein, the product of *glcA*, is produced in the transformants. An activity assay was used to determine whether glycerol dehydrogenase activity remained in the transformants. Activity in glycerol dehydrogenase bands on native gels was determined by coupling the conversion of glycerol + NAD(+) → dihydroxyacetone + NADH to the conversion of a tetrazolium dye, MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-

diphenyltetrazolium bromide] to a deeply colored formazan, with phenazine methosulfate as mediator. Glycerol dehydrogenase also requires the presence of 30 mM ammonium sulfate and 100 mM Tris, pH 9 (C.-T. Tang, et al. (1997) *J. Bacteriol.* 140:182). Of 8 transformants analyzed, 6 were determined to be 5 *gldA* knockouts. *E. coli* MSP33.6 has been deposited with ATCC under the terms of the Budapest Treaty on 24 November 1997.

EXAMPLE 7

CONSTRUCTION OF *E. COLI* STRAIN WITH *GLPK*
AND *GLDA* GENE KNOCKOUTS

10 A 1.6 kb DNA fragment containing the *gldA* gene, and including 228 bps of DNA upstream of the translational start codon and 220 bps of DNA downstream of the translational stop codon was isolated from *E. coli* by PCR using primers SEQ ID NO:42 and SEQ ID NO:43, which incorporate terminal Sph1 and Xba1 sites, respectively, and cloned between the Sph1 and Xba1 sites 15 of pUC18, to generate pQN2. pQN2 was cut at the unique Sal1 and Nco1 sites within the *gldA* gene, the ends flushed with Klenow and religated, resulting in a 109 bp deletion in the middle of *gldA* and regeneration of a unique Sal1 site, to generate pQN4. A 1.2 kb DNA fragment containing the gene conferring kanamycin resistance (kan), and flanked by loxP sites was isolated from 20 pLoxKan2 (Genecor International, Palo Alto, Calif.) as a Stu1/Xho1 fragment, the ends flushed with Klenow, and subcloned into pQN4 at the Sal1 site after flushing with Klenow, to generate pQN8. The Sph1/Xba1 fragment from pQN8 containing the kan-interrupted *gldA* was subcloned between the Sph1 and Xba1 sites of pGP704, using *E. coli* SY327 as host, to generate pQN9. *E. coli* RJF10 25 (see EXAMPLE 5) was transformed with pQN9 and selected on kan. Since the pGP704 backbone cannot replicate in most *E. coli* hosts, transformants arise by integration of the plasmid, (or portions of it) into the chromosome. Double crossover integration events are determined by identifying those transformants which are kan resistant and ampicillin sensitive. PCR and Southern blot 30 analyses are used to determine the nature of the chromosomal integration events in the transformants. Western blot analysis is used to determine whether glycerol dehydrogenase, the product of *gldA*, is produced in the transformants. Activity assays are used to determine whether glycerol dehydrogenase activity remains in the transformants. The kan marker is removed from the chromosome 35 using the Cre-producing plasmid pJW168, as described in EXAMPLE 5.

EXAMPLE 8

CONSUMPTION OF GLYCEROL PRODUCED FROM D-GLUCOSE BY RECOMBINANT *E. COLI* CONTAINING BOTH GPP2 AND DAR1 WITH AND WITHOUT GLYCEROL KINASE (GLPK) ACTIVITY

5 EXAMPLE 8 illustrates the consumption of glycerol by the recombinant *E. coli* FM5/pAH48 and RJF10/pAH48. The strains FM5/pAH48 and RJF10/pAH48 were constructed as described above in the GENERAL METHODS.

Pre-Culture

10 FM5/pAH48 and RJF10/pAH48 were pre-cultured for seeding a fermenter in the same medium used for fermentation, or in LB supplemented with 1% glucose. Either carbenicillin or ampicillin were used (100 mg/L) for plasmid maintenance. The medium for fermentation is as described in EXAMPLE 4.

15 Cultures were started from frozen stocks (15% glycerol as cryoprotectant) in 600 mL medium in a 2-L Erlenmeyer flask, grown at 30 °C in a shaker at 250 rpm for approximately 12 h, and used to seed the fermenter.

Fermentation growth

20 A 15-L stirred tank fermenter with 5-7 L initial volume was prepared as described in EXAMPLE 4. Either carbenicillin or ampicillin were used (100 mg/L) for plasmid maintenance.

Environmental Conditions to Evaluate Glycerol Kinase (GlpK) Activity

25 The temperature was controlled at 30 °C and the air flow rate controlled at 6 standard liters per minute. Back pressure was controlled at 0.5 bar. Dissolved oxygen tension was controlled at 10% by stirring. Aqueous ammonia was used to control pH at 6.7. The glucose feed (60% glucose) rate was controlled to maintain excess glucose until glycerol had accumulated to at least 25 g/L. Glucose was then depleted, resulting in the net metabolism of glycerol. Table 2 shows the resulting conversion of glycerol.

30

Table 2
Conversion of glycerol by FM5/pAH48 (wt) and RJF10/pAH48 (glpK)

Strain	number of examples	rate of glycerol consumption g/OD/hr
FM5/pAH48	2	0.095 ± 0.015
RJF10/pAH48	3	0.021 ± 0.011

As is seen by the data in Table 2, the rate of glycerol consumption decreases about 4-5 fold where endogenous glycerol kinase activity is eliminated.

Environmental Conditions to Evaluate Glycerol Dehydrogenase (GldA) Activity

5 The temperature was controlled at 30 °C and the air flow rate controlled at 6 standard liters per minute. Back pressure was controlled at 0.5 bar. Dissolved oxygen tension was controlled at 10% by stirring. Aqueous ammonia was used to control pH at 6.7. In the first fermentation, glucose was kept in excess for the duration of the fermentation. The second fermentation was
10 operated with no residual glucose after the first 25 hours. Samples over time from the two fermentations were taken for evaluation of GlpK and GldA activities. Table 3 summarizes RJF10/pAH48 fermentations that show the effects of GldA on selectivity for glycerol.

Table 3
GldA and GlpK activities from two RJF10/pAH48 fermentations

Fermentation	Time (hrs)	GldA	GlpK	Overall selectivity (g/g)
1	25	-	-	42%
	46	-	-	49%
	61	+	-	54%
2	25	+	-	41%
	46	++	-	14%
	61	++	-	12%

15 As is seen by the data in Table 3, the presence of glycerol dehydrogenase (GldA) activity is linked to the conversion of glycerol under glucose-limited conditions; thus, it is anticipated that eliminating glycerol dehydrogenase activity will reduce glycerol conversion.

20

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(B) COMPUTER: IBM PC COMPATIBLE
(C) OPERATING SYSTEM: MICROSOFT WINDOWS 95
(D) SOFTWARE: MICROSOFT WORD VERSION 7.0A

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: 60/030602
(B) FILING DATE: NOVEMBER 13, 1996
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: FLOYD, LINDA AXAMETHY
(B) REGISTRATION NUMBER: 33,692
(C) REFERENCE/DOCKET NUMBER: CR-9981-B

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: 302-892-8112
(B) TELEFAX: 302-773-0164

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1380 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CTTTAATTTT	CTTTTATCTT	ACTCTCCACT	ATAAGACATC	AAGAAACAAT	TGTATATTGT	60
ACACCCCCCC	CCTCCACAAA	CACAAATATT	GATAATATAA	AGATGTCTGC	TGCTGCTGAT	120
AGATTAAACT	TAACTTCCGG	CCACTTGAAT	GCTGGTAGAA	AGAGAAGTTC	CTCTTCTGTT	180
TCTTGAGG	CTGCCGAAAA	GCCTTCAAG	GTAACTGTGA	TTGGATCTGG	TAACTGGGT	240
ACTACTATTG	CCAAGGTGGT	TGCCGAAAAT	TGTAAGGGAT	ACCCAGAAGT	TTTCGCTCCA	300
ATAGTACAAA	TGTGGGTGTT	CGAAGAAGAG	ATCAATGGTG	AAAAATTGAC	TGAAATCATA	360
AATACTAGAC	ATCAAAACGT	GAAATACTTG	CCTGGCATCA	CTCTACCCGA	CAATTGGTT	420
GCTAATCCAG	ACTTGATTGA	TTCAGTCAAG	GATGTCGACA	TCATCGTTT	CAACATTCCA	480
CATCAATTTC	TGCCCGTAT	CTGTAGCCAA	TTGAAAGGTC	ATGTTGATTC	ACACGTCAGA	540
GCTATCTCCT	GTCTAAAGGG	TTTGAAAGTT	GGTGCTAAAG	GTGTCCAATT	GCTATCCTCT	600
TACATCACTG	AGGAACACTAG	TATTCAATGT	GGTGCCTAT	CTGGTGCCTAA	CATTGCCACC	660
GAAGTCGCTC	AAGAACACTG	GTCGAAACA	ACAGTTGCTT	ACCACATTCC	AAAGGATTTC	720
AGAGGCGAGG	GCAAGGACGT	CGACCATAAAG	GTCTCAAAGG	CCTTGTCTCA	CAGACCTTAC	780
TTCCACGTTA	GTGTCATCGA	AGATGTTGCT	GGTATCTCCA	TCTGTGGTGC	TTTGAAGAAC	840
GTTGTTGCCT	TAGGGTGTGG	TTTCGTCGAA	GGTCTAGGCT	GGGGTAAACAA	CGCTTCTGCT	900
GCCATCCAAA	GAGTCGGTTT	GGGTGAGATC	ATCAGATTG	GTCAAATGTT	TTTCCCAGAA	960
TCTAGAGAAC	AAACATACTA	CCAAGAGTCT	GCTGGTGTG	CTGATTGAT	CACCACCTGC	1020
GCTGGTGGTA	GAAACGTCAA	GGTGCTAGG	CTAATGGCTA	CTTCTGGTAA	GGACGCCCTGG	1080
GAATGTGAAA	AGGAGTTGTT	GAATGGCCAA	TCCGCTCAAG	GTAAATTAC	CTGCAAAGAA	1140
GTTCACGAAT	GGTTGGAAC	ATGTGGCTCT	GTGGAAGACT	TCCCATTATT	TGAAGCCGTA	1200
TACCAAATCG	TTTACAACAA	CTACCCAATG	AAGAACCTGC	CGGACATGAT	TGAAGAATTA	1260
GATCTACATG	AAGATTAGAT	TTATTGGAGA	AAGATAACAT	ATCATACTTC	CCCCACTTTT	1320
TTCGAGGCTC	TTCTATATCA	TATTCAAA	TTAGCATTAT	GTCATTTCTC	ATAACTACTT	1380

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2946 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

GAATTCGAGC	CTGAAAGTGCT	GATTACCTTC	AGGTAGACTT	CATCTTGACC	CATCAACCCC	60
AGCGTCAATC	CTGCAAATAC	ACCACCCAGC	AGCAGTAGGA	TGATAGAGAT	AATATAGTAC	120
GTGGTAACGC	TTGCCCTCATC	ACCTACGCTA	TGGCCGGAAT	CGGCAACATC	CCTAGAATTG	180
AGTACGTGTG	ATCCGGATAA	CAACGGCAGT	GAATATATCT	TCGGTATCGT	AAAGATGTGA	240
TATAAGATGTA	TGTATACCCA	ATGAGGAGCG	CCTGATCGTG	ACCTAGACCT	TAGTGGCAA	300
AACGACATAT	CTATTATAGT	GGGGAGAGTT	TCGTGCAAAT	AACAGACGCA	GCAGCAAGTA	360
ACTGTGACGA	TATCAACTCT	TTTTTATTAA	TGTAATAAGC	AAACAAGCAC	GAATGGGGAA	420
AGCCTATGTG	CAATCACCAA	GGTCGTCCT	TTTTTCCCAT	TTGCTAATT	AGAATTAAA	480
GAAACCAAAA	GAATGAAGAA	AGAAAACAAA	TACTAGCCCT	AAACCTGACT	TCGTTCTAT	540
GATAATACCC	TGCTTAAATG	AAACGGTATGC	CCTAGGGTAT	ATCTCACTCT	GTACGTTACA	600
AACTCCGGTT	ATTTTATCGG	AAACATCCGAG	CACCCGCGCC	TTCCTCAACC	CAGGCACCGC	660
CCCAGGTAAC	CGTGCACGAT	GAGCTAATCC	TGAGCCATCA	CCCACCCAC	CCGTTGATGA	720
CAGCAATTG	GGAGGGCGAA	AATAAAACTG	GAGCAAGGAA	TTACCATCAC	CGTCACCAC	780
ACCATCATAT	CGCCTTAGCC	TCTAGCCATA	GCCATCATGC	AAGCGTGTAT	CTTCTAAGAT	840
TCAGTCATCA	TCATTACCGA	GTTTGTTC	CTTCACATGA	TGAAGAAGGT	TTGAGTATGC	900
TCGAAAACAAT	AAGACGACGA	TGGCTCTGCC	ATTGGTTATA	TTACGCTTT	CGGGCGAGGT	960
GGCGATGGGT	TGCTGAGGGG	AAAGAGTGT	AGCTTACGGA	CCTATTGCCA	TTGTTATTCC	1020
GATTAATCTA	TTGGTCAGCA	GCTCTCTCT	ACCCGTCTAT	TCTAGTATTT	TTTTTTTTT	1080
TTTTGGTTT	TACTTTTTT	TCTTCTTGCC	TTTTTTCTT	GTACTTTT	TTCTAGTTT	1140
TTTTCTTC	ACTAAGCTT	TTCCCTGATT	TATCCTGGG	TTCTTCTTC	TACTCCTTA	1200
GATTTTTTTT	TTATATATTA	ATTTTAAGT	TTATGTATTT	TGGTAGATTC	AATTCTCTT	1260
CCCTTCCCTT	TCCTTCGCT	CCCCCTCCCTT	ATCAATGCTT	GCTGTCAGAA	GATTAACAAG	1320
ATACACATTC	CTTAAGCGAA	CGCATCCGGT	GTATATACT	CGTCGTGCAT	ATAAAATTT	1380

GCCTTCAAGA TCTACTTTCC TAAGAAGATC ATTATTACAA ACACAAC TGC ACTCAAAGAT 1440
 GACTGCTCAT ACTAATATCA AACAGCACAA ACAC TGT CAT GAGGACCATC CTATCAGAAG 1500
 ATCGGACTCT GCCGTGTCAA TTGTACATT GAAACGTGCG CCCTTCAAGG TTACAGTGAT 1560
 TGGTTCTGGT AACTGGGGGA CCACCATCGC CAAAGTCATT GCGGAAAACA CAGAATTGCA 1620
 TTCCCATATC TTGAGGACAG AGGTGAGAAT GTGGGTTTT GATGAAAAGA TCGGCGACGA 1680
 AAATCTGACG GATATCATAA ATACAAGACA CCAGAACGTT AAATATCTAC CCAATATTGA 1740
 CCTGCCCAT AATCTAGTGG CCGATCCTGA TCTTTACAC TCCATCAAGG GTGCTGACAT 1800
 CCTTGTTC AACATCCCTC ATCAATT TTT ACCAACATA GTCAAACAAAT TGCAAGGCCA 1860
 CGTGGCCCT CATGTAAGGG CCATCTCGT TCTAAAAGGG TTCGAGTTGG GCTCCAAGGG 1920
 TGTGCAATTG CTATCCTCCT ATGTTACTGA TGAGTTAGGA ATCCAATGTG GCGCACTATC 1980
 TGGTGCAACAC TTGGCACCGG AAGTGGCCAA GGAGCATTGG TCCGAAACCA CC GTGGCTTA 2040
 CCAACTACCA AAGGATTATC AAGGTGATGG CAAGGATGTA GATCATAAGA TTTTGAAATT 2100
 GCTGTCCAC AGACCTACT TCCACGTCAA TGTATCGAT GATGTTGCTG GTATATCCAT 2160
 TGCCGGTGCC TTGAAGAACG TCGTGGCACT TGCATGTGGT TTCGTAGAAG GTATGGGATG 2220
 GGGTAACAAAT GCCTCCGAG CCATTCAAAG GCTGGGTTTA GGTGAAATT TCAAGTTCCG 2280
 TAGAATGTTT TTCCCAGAAT CCAAAGTCGA GACCTACTAT CAAGAATCCG CTGGTGTG 2340
 AGATCTGATC ACCACCTGCT CAGGGCGTAG AAACGTCAAG GTGCCACAT ACATGGCCAA 2400
 GACCGGTAAG TCAGCCTTGG AAGCAGAAAA GGAATTGCTT AACGGTCAAT CCGCCCAAGG 2460
 GATAATCACA TGCAGAGAACG TTCAAGAGTG GCTACAAACA TGTGAGTTGA CCCAAGAATT 2520
 CCAAATTATT CGAGGCAGTC TACCA GATAG TCTACAAACAA CGTCCGCATG GAAGACCTAC 2580
 CGGAGATGAT TGAAGAGCTA GACATCGATG AGCAATAGAC ACTCTCCCCC CCCCTCCCC 2640
 TCTGATCTTT CCTGTTGCCT CTTTTCCCC CAACCAATT ATCATTATAC ACAAGTTCTA 2700
 CAACTACTAC TAGTAACATT ACTACAGTTA TTATAATT TCTATTCTT TTTCTTTAAG 2760
 AATCTATCAT TAACGTTAAT TTCTATATAT ACATAACTAC CATTATACAC GCTATTATCG 2820
 TTTACATATC ACATCACCGT TAATGAAAGA TAGCACACCC TGTACACTAA CACAATTAAA 2880
 TAATGCCAT AACCTTTCT GTTATCTATA GCCCTTAAAG CTGTTCTTC GAGCTTTCA 2940
 CTGCAG 2946

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 3178 base pairs

(B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CTGCAGAACT TCGCTGCTC TGTGCCCATC CTCGCGTTA GAAAAGAGCT GAATTGTTTC	60
ATGCGCAAGG GCATCAGCGA GTGACCAATA ATCACTGCAC TAATCCTTT TTAGCAACAC	120
ATACCTTATAT ACAGCACCAG ACCTTATGTC TTTCTCTGC TCCGATACGT TATCCCACCC	180
AACTTTATT TCAGTTTTG CAGGGGAAAT TTCACAAACCC CGCACGCTAA AAATCGTATT	240
TAAACTTAAA AGAGAACAGC CACAAATAGG GAACTTTGGT CTAAACGAAG GACTCTCCCT	300
CCCTTATCTT GACCGTGTCA TTGGCCATCAC TGCTACAAAGA CTAAATACGT ACTAATATAT	360
GTTCCTCGTA ACGAGAAGAA GAGCTGCCGG TGCAGCTGCT GCCATGGCCA CAGCCACGGG	420
GACGCTGTAC TGGATGACTA GCCAAGGTGA TAGGCCGTAA GTGCACAATG ACCCGAGCTA	480
CATGGTGCAA TTCCCCACCG CCGCTCCACC GGCAGGTCTC TAGACGAGAC CTGCTGGACC	540
GTCTGGACAA GACGCATCAA TTGACGTGT TGATCATCGG TGCGGGGGCC ACGGGGACAG	600
GATGTGCCCT AGATGCTGCG ACCAGGGGAC TCAATGTGGC CCTTGTGAA AAGGGGGATT	660
TTGCCTCGGG AACGTCGTCC AAATCTACCA AGATGATTCA CGGTGGGGTG CGGTACTTAG	720
AGAAGGCCTT CTGGGAGTTC TCCAAGGCAC AACTGGATCT GGTCATCGAG GCACCTCAACG	780
AGCGTAAACA TCTTATCAC ACTGCCCTC ACCTGTGCAC GGTGCTACCA ATTCTGATCC	840
CCATGTACAG CACCTGGCAG GTCCCGTACA TCTATATGGG CTGTAATTC TACGATTCT	900
TTGGCGGTTC CCAAAACTTG AAAAATCAT ACCTACTGTC CAAATCCGCC ACCGTGGAGA	960
AGGCTCCCAT GCTTACCAACA GACAATTAA AGGCCTCGCT TGTGTACCAT GATGGGTCT	1020
TTAACGACTC GCGTTGAAC GCCACTTTCG CCATCACCGG TGTGGAGAAC GGCCTACCG	1080
TCTTGATCTA TGTCGAGGTA CAAAAATTGA TCAAAAGACCC AACTTCTGGT AAGGTTATCG	1140
GTGCCGAGGC CGGGGACGTT GAGACTAATG AGCTTGTCAAG AATCAACGCT AAATGTGTGG	1200
TCAATGCCAC GGGCCCACAT AGTGACGCCA TTTGCAAAT GGACCGCAAC CCATCCGGTC	1260
TGCCGGACTC CCCGCTAACAC GACAACCTCA AGATCAAGTC GACTTCAAT CAAATCTCCG	1320
TCATGGACCC GAAAATGGTC ATCCCATCTA TTGGCGTTCA CATCGTATTG CCCTCTTTT	1380
ACTCCCCGAA GGATATGGGT TTGGTGGACG TCAGAACCTC TGATGGCAGA GTGATGTTCT	1440
TTTACCTTG GCAGGGAAA GTCCTTGCCG GCACCCACAGA CATCCCACTA AAGCAAGTCC	1500

CAGAAAACCC	TATGCCTACA	GAGGCTGATA	TTCAAGATAT	CTTGAAGAGAA	CTACAGCACT	1560
ATATCGAATT	CCCCGTGAAA	AGAGAAGACG	TGCTAAGTGC	ATGGGCTGGT	GTCAGACCTT	1620
TGGTCAGAGA	TCCACGTACA	ATCCCCGCAG	ACGGGAAGAA	GGGCTCTGCC	ACTCAGGGCG	1680
TGGTAAGATC	CCACTTCTTG	TTCACTCGG	ATAATGGCCT	AATTACTATT	GCAGGTGGTA	1740
AATGGACTAC	TTACAGACAA	ATGGCTGAGG	AAACAGTCGA	CAAAGTTGTC	GAAGTTGGCG	1800
GATTCCACAA	CCTGAAACCT	TGTCACACAA	GAGATATTAA	GCTTGCTGGT	GCAGAGAAAT	1860
GGACGCAAA	CTATGTGGCT	TTATTGGCTC	AAAACTACCA	TTTATCATCA	AAAATGTCCA	1920
ACTACTTGGT	TCAAAACTAC	GGAACCCGTT	CCTCTATCAT	TTGCGAATT	TTCAGAAGAAAT	1980
CCATGGAAAA	AAACTGCGCT	TTGTCCTTAG	CCGACAAGGA	AAATAACGTA	ATCTACTCTA	2040
GCGAGGGAGAA	CAACTGGTC	AAAGAGGATA	CTTCAGATA	TCCATTACACA	ATCGGTGAGT	2100
TAAAGTATT	CATGCAGTAC	GAATATTGTA	GAACTCCCTT	GGACTTCCTT	TTAAGAAGAA	2160
CAAGATTCGC	CTTCTTGGAC	GCCAAGGAAG	CTTGTGATGC	CGTGCATGCC	ACCGTCAAAG	2220
TTATGGGTGA	TGAGTTCAAT	TGGTCGGAGA	AAAAGAGGCA	GTGGGAACTT	AAAAAAACTG	2280
TGAACTTCAT	CCAAGGACGT	TTCGGTGTCT	AAATCGATCA	TGATAGTTAA	GGGTGACAAA	2340
GATAACATTC	ACAAGAGTAA	TAATAATGGT	AATGATGATA	ATAATAATAA	TGATAGTAAT	2400
AACAATAATA	ATAATGGTGG	TAATGGCAAT	GAAATCGCTA	TTATTACCTA	TTTTCCCTAA	2460
TGGAAGAGTT	AAAGTAAACT	AAAAAAACTA	CAAAATATA	TGAAGAAAAAA	AAAAAAAAGA	2520
GGTAATAGAC	TCTACTACTA	CAATTGATCT	TCAAATTATG	ACCTTCCTAG	TGTTTATATT	2580
CTATTCCAA	TACATAATAT	AATCTATATA	ATCATTGCTG	GTAGACTTCC	GTTTAATAT	2640
CGTTTAATT	ATCCCTTTA	TCTCTAGTCT	AGTTTATCA	AAAATATAG	AAACACTAAA	2700
TAATATTCTT	CAAACGGTCC	TGGTGATAC	GCAATACATA	TTTATGGTGC	AAAAAAAAGA	2760
ATGGAAAATT	TTGCTAGTCA	TAACCCTTT	CATAAAACAA	TACGTAGACA	TCGCTACTTG	2820
AAATTTCAA	GTTTTATCA	GATCCATGTT	TCCTATCTGC	CTTGACAAACC	TCATCGTCGA	2880
AATAGTACCA	TTTAGAACGC	CCAATATTCA	CATTGTGTTC	AAGGTCTTTA	TTCACCAGTG	2940
ACGTGTAATG	GCCATGATTA	ATGTGCCTGT	ATGGTTAAC	ACTCCAAATA	GCTTATATTT	3000
CATAGTGTCA	TTGTTTTCA	ATATAATGTT	TAGTATCAAT	GGATATGTTA	CGACGGTGT	3060
ATTTTCTTG	GTCAAATCGT	AATAAAATCT	CGATAATGG	ATGACTAAGA	TTTTGGTAA	3120
AGTTACAAA	TTTATCGTTT	TCACTGTTGT	CAATTTTTG	TTCTTGTAA	CACTCGAG	3178

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 816 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

ATGAAACGTT TCAATGTTT AAAATATATC AGAACACAA AAGCAAATAT ACAAACATC	60
GCAATGCCTT TGACCACAAA ACCTTATCT TTGAAAATCA ACGCCGCTCT ATTGATGTT	120
GACGGTACCA TCATCATCTC TCAACCAGCC ATTGCTGCTT TCTGGAGAGA TTTCGGTAA	180
GACAAGCCTT ACTTCGATGC CGAACACGTT ATTACATCT CTCACGGTTG GAGAACTTAC	240
GATGCCATTG CCAAGTCGCG TCCAGACTTT GCTGATGAAG AATACGTTAA CAAGCTAGAA	300
GGTGAAATCC CAGAAAAGTA CGGTGAACAC TCCATCGAAG TTCCAGGTGC TGTCAAGTTG	360
TGTAATGCTT TGAACGCCTT GCCAAAGGAA AAATGGGCTG TCGCCACCTC TGGTACCGGT	420
GACATGGCCA AGAAATGGTT CGACATTTG AAGATCAAGA GACCAGAATA CTTCATCACC	480
GCCAATGATG TCAAGCAAGG TAAGCCTCAC CCAGAACCAT ACTTAAAGGG TAGAAACGGT	540
TTGGGTTCC CAATTAATGA ACAAGACCCA TCCAAATCTA AGGTTGTTGT CTTGAAGAC	600
GCACCAAGCTG GTATTGCTGC TGGTAAGGCT GCTGGCTGTA AAATCGTTGG TATTGCTACC	660
ACTTCGATT TGGACTTCTT GAAGGAAAAG GGTGTTGACA TCATTGTCAA GAACCACGAA	720
TCTATCAGAG TCGGTGAATA CAACGCTGAA ACCGATGAAG TCGAATTGAT CTTGATGAC	780
TACTTATAACG CTAAGGATGA CTTGTTGAAA TGGTAA	816

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 753 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

ATGGGATTGA CTACTAAACC TCTATCTTG AAAGTTAACG CCGCTTGTT CGACGTCGAC	60
GGTACCATTA TCATCTCTCA ACCAGCCATT GCTGCATTCT GGAGGGATTT CGGTAAGGAC	120
AAACCTTATT TCGATGCTGA ACACGTTATC CAAGTCTCGG ATGGTTGGAG AACGTTGAT	180

GCCATTGCTA	AGTCGCTCC	AGACTTTGCC	AATGAAGAGT	ATGTTAACAA	ATTAGAAGCT	240
GAAATTCCGG	TCAAGTACGG	TGAAAAATCC	ATTGAAGTCC	CAGGTGCAGT	TAAGCTGTGC	300
AACGCTTGA	ACGCTCTACC	AAAAGAGAAA	TGGGCTGTGG	CAACTCCGG	TACCCGTGAT	360
ATGGCACAAA	AATGGTTCGA	GCATCTGGGA	ATCAGGAGAC	CAAAGTACTT	CATTACCGCT	420
AATGATGTCA	AACAGGGTAA	GCCTCATCCA	GAACCATATC	TGAAGGGCAG	GAATGGCTTA	480
GGATATCCGA	TCAATGAGCA	AGACCCTTCC	AAATCTAAGG	TAGTAGTATT	TGAAGACGCT	540
CCAGCAGGTA	TTGCCGCCGG	AAAAGCCGCC	GGTTGTAAGA	TCATTGGTAT	TGCCACTACT	600
TCAGACTTGG	ACTTCCTAAA	GGAAAAAGGC	TGTGACATCA	TTGTCAAAAA	CCACGAATCC	660
ATCAGAGTTG	GGGGCTACAA	TGCCGAAACA	GACGAAGTTG	AATTCACTTT	TGACGACTAC	720
TTATATGCTA	AGGACGATCT	GTTGAAATGG	TA			753

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2520 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

TGTATTGGCC	ACGATAACCA	CCCTTTGTAT	ACTGTTTTTG	TTTTTCACAT	GTTAAATAAC	60
GACTTTTATT	AAACAACGTA	TGTAAAAACA	TAACAAGAAT	CTACCCATAC	AGGCCATTTC	120
GTAATTCTTC	TCTTCTTAATT	GGAGTAAAC	CATCAATTAA	AGGGTGTGGA	GTAGCATAGT	180
GAGGGGCTGA	CTGCATTGAC	AAAAAAATTG	AAAAAAAAAA	AGGAAAAGGA	AAGGAAAAAA	240
AGACAGCCAA	GACTTTAGA	ACGGATAAGG	TGTATAAAA	TGTGGGGGGA	TGCCTGTTCT	300
CGAACCATAT	AAAATATACC	ATGTGGTTG	AGTTGTGGCC	GGAACATATAC	AAATAGTTAT	360
ATGTTCCCT	CTCTCTTCCG	ACTTGTAGTA	TTCTCCAAAC	GTACATATT	CCGATCAAGC	420
CAGCGCCTT	ACACTAGTTT	AAAACAAGAA	CAGAGCCGTA	TGTCCAAAT	AATGGAAGAT	480
TTACGAAGTG	ACTACGTCCC	GCTTATCGCC	AGTATTGATG	TAGGAACGAC	CTCATCCAGA	540
TGCATTCTGT	TCAACAGATG	GGGCCAGGAC	GTTCAAAAC	ACCAAATTGA	ATATTCAACT	600
TCAGCATCGA	AGGGCAAGAT	TGGGGTGTCT	GGCTTAAGGA	GACCCCTCTAC	AGCCCCAGCT	660
CGTGAACAC	CAAACGCCGG	TGACATCAAA	ACCAGCGGAA	AGCCCATCTT	TTCTGCAGAA	720
GGCTATGCCA	TTCAAGAAC	CAAATTCTA	AAAATCGAGG	AATTGGACTT	GGACTTCCAT	780

AACGAACCCA	CGTTGAAGTT	CCCCAAACCG	GGTTGGGTTG	AGTGCCATCC	GCAGAAATTA	840
CTGGTGAACG	TCGTCCTAATG	CCTTGCTCA	AGTTTGCTCT	CTCTGCAGAC	TATCACACGC	900
GAACGTGTAG	CAAACGGTCT	CCCACCTTAC	AAGGTAATAT	GCATGGGTAT	AGCAAACATG	960
AGAGAAACCA	CAATTCTGTG	GTCCCCGCCGC	ACAGGAAAAC	CAATTGTTAA	CTACGGTATT	1020
GTGGAAACG	ACACCAAGAAC	GATCAAATC	GTTAGAGACA	AATGGCAAAA	CACTAGCGTC	1080
GATAGGCAAC	TGAGCTTAG	ACAGAAGACT	GGATTGCCAT	TGCTCTCCAC	GTATTTCTCC	1140
TGTTCCAAGC	TGCGCTGGTT	CCTCGACAAT	GAGCCTCTGT	GTACCAAGGC	GTATGAGGAG	1200
AACGACCTGA	TGTCGGCAC	TGTGGACACA	TGGCTGATTT	ACCAATTAAC	TAACAAACAA	1260
GCCTTCTTT	CTGACGTAAC	CAACGCTTCC	AGAACTGGAT	TTATGAAACCT	CTCCACTTTA	1320
AAGTACGACA	ACGAGTTGCT	GGAAATTGG	GGTATTGACA	AGAACCTGAT	TCACATGCC	1380
GAAATTGTGT	CCTCATCTCA	ATACTACGGT	GACTTTGGCA	TTCCTGATTG	GATAATGGAA	1440
AAGGTACACG	ATTGCCAAA	AAACAGTACTG	CGAGATCTAG	TCAAGAGAAA	CCTGCCATA	1500
CAGGGCTGTC	TGGGCGACCA	AAAGCGCATCC	ATGGTGGGGC	AACTCGCTTA	CAAACCCGGT	1560
GCTGAAAAT	GTACTTATGG	TACCGGTTGC	TTTTTACTGT	ACAATACGGG	GACCAAAAAAA	1620
TTGATCTCCC	AACATGGCGC	ACTGACGACT	CTAGCATTTC	GGTCCCCACA	TTTGCAAGAG	1680
TACGGTGGCC	AAAAACCCAGA	ATTGAGCAAG	CCACATTTTG	CATTAGAGGG	TTCCGTCGCT	1740
GTGGCTGGTG	CTGTGGTCCA	ATGGCTACGT	GATAATTAC	GATTGATCGA	TAATACAGAG	1800
GATGTCGGAC	CGATTGCATC	TACGGTTCC	GATTCTGGTG	GCGTAGTTT	CGTCCCCGCA	1860
TTTAGTGGCC	TATTGCTCC	CTATTGGAC	CCAGATGCCA	GAGCCACCAT	AATGGGGATG	1920
TCTCAATTCA	CTACTGCC	CCACATGCC	AGAGCTGCCG	TGGAAGGTGT	TTGCTTTCAA	1980
GCCAGGGCTA	TCTTGAAGGC	AATGAGTTCT	GACCGCTTTC	GTGAAGGTT	CAAAGACAGG	2040
GACTTTTTAG	AGGAAATTTC	CGACGTACA	TATGAAAAGT	CGCCCCGTC	GGTTCTGGCA	2100
GTGGATGGCG	GGATGTCGAG	GTCTAATGAA	GTCTGCAA	TTCAAGCCGA	TATCCTAGGT	2160
CCCTGTGTCA	AAGTCAGAAG	GTCTCCGACA	GCGGAATGTA	CCGCATTGGG	GGCAGCCATT	2220
GCAGCCAATA	TGGCTTCAA	GGATGTAAC	GAGCGCCCAT	TATGAAAGGA	CCTACACGAT	2280
GTAAAGAAAT	GGGTCTTTA	CAATGGAATG	GAGAAAAACG	AACAAATATC	ACCAAGGGCT	2340
CATCCAAACC	TTAAGATATT	CAGAAGTGAA	TCCGACGATG	CTGAAAGGAG	AAAGCATTGG	2400
AGTATTGGG	AAAGTGGCGT	GGAAAGATCC	AAAGGTTGGC	TGAAGGACAT	AGAAGGTGAA	2460
CACCAACAGG	TTCTAGAAAA	CTTCCAATAA	CAACATAAT	AATTCTATT	AAACATGTAA	2520

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 391 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Met	Ser	Ala	Ala	Ala	Asp	Arg	Leu	Asn	Leu	Thr	Ser	Gly	His	Leu	Asn
1							5			10				15	
Ala Gly Arg Lys Arg Ser Ser Ser Val Ser Leu Lys Ala Ala Glu															
							20			25			30		
Lys Pro Phe Lys Val Thr Val Ile Gly Ser Gly Asn Trp Gly Thr Thr															
							35			40			45		
Ile Ala Lys Val Val Ala Glu Asn Cys Lys Gly Tyr Pro Glu Val Phe															
							50			55			60		
Ala Pro Ile Val Gln Met Trp Val Phe Glu Glu Glu Ile Asn Gly Glu															
							65			70			75		80
Lys Leu Thr Glu Ile Ile Asn Thr Arg His Gln Asn Val Lys Tyr Leu															
							85			90			95		
Pro Gly Ile Thr Leu Pro Asp Asn Leu Val Ala Asn Pro Asp Leu Ile															
							100			105			110		
Asp Ser Val Lys Asp Val Asp Ile Ile Val Phe Asn Ile Pro His Gln															
							115			120			125		
Phe Leu Pro Arg Ile Cys Ser Gln Leu Lys Gly His Val Asp Ser His															
							130			135			140		
Val Arg Ala Ile Ser Cys Leu Lys Gly Phe Glu Val Gly Ala Lys Gly															
							145			150			155		160
Val Gln Leu Leu Ser Ser Tyr Ile Thr Glu Glu Leu Gly Ile Gln Cys															
							165			170			175		
Gly Ala Leu Ser Gly Ala Asn Ile Ala Thr Glu Val Ala Gln Glu His															
							180			185			190		
Trp Ser Glu Thr Thr Val Ala Tyr His Ile Pro Lys Asp Phe Arg Gly															
							195			200			205		
Glu Gly Lys Asp Val Asp His Lys Val Leu Lys Ala Leu Phe His Arg															
							210			215			220		
Pro Tyr Phe His Val Ser Val Ile Glu Asp Val Ala Gly Ile Ser Ile															
							225			230			235		240

Cys Gly Ala Leu Lys Asn Val Val Ala Leu Gly Cys Gly Phe Val Glu
 245 250 255
 Gly Leu Gly Trp Gly Asn Asn Ala Ser Ala Ala Ile Gln Arg Val Gly
 260 265 270
 Leu Gly Glu Ile Ile Arg Phe Gly Gln Met Phe Phe Pro Glu Ser Arg
 275 280 285
 Glu Glu Thr Tyr Tyr Gln Glu Ser Ala Gly Val Ala Asp Leu Ile Thr
 290 295 300
 Thr Cys Ala Gly Gly Arg Asn Val Lys Val Ala Arg Leu Met Ala Thr
 305 310 315 320
 Ser Gly Lys Asp Ala Trp Glu Cys Glu Lys Glu Leu Leu Asn Gly Gln
 325 330 335
 Ser Ala Gln Gly Leu Ile Thr Cys Lys Glu Val His Glu Trp Leu Glu
 340 345 350
 Thr Cys Gly Ser Val Glu Asp Phe Pro Leu Phe Glu Ala Val Tyr Gln
 355 360 365
 Ile Val Tyr Asn Asn Tyr Pro Met Lys Asn Leu Pro Asp Met Ile Glu
 370 375 380
 Glu Leu Asp Leu His Glu Asp
 385 390

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 384 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Thr Ala His Thr Asn Ile Lys Gln His Lys His Cys His Glu Asp
 1 5 10 15
 His Pro Ile Arg Arg Ser Asp Ser Ala Val Ser Ile Val His Leu Lys
 20 25 30
 Arg Ala Pro Phe Lys Val Thr Val Ile Gly Ser Gly Asn Trp Gly Thr
 35 40 45
 Thr Ile Ala Lys Val Ile Ala Glu Asn Thr Glu Leu His Ser His Ile
 50 55 60
 Phe Glu Pro Glu Val Arg Met Trp Val Phe Asp Glu Lys Ile Gly Asp
 65 70 75 80

Glu Asn Leu Thr Asp Ile Ile Asn Thr Arg His Gln Asn Val Lys Tyr
85 90 95

Leu Pro Asn Ile Asp Leu Pro His Asn Leu Val Ala Asp Pro Asp Leu
100 105 110

Leu His Ser Ile Lys Gly Ala Asp Ile Leu Val Phe Asn Ile Pro His
115 120 125

Gln Phe Leu Pro Asn Ile Val Lys Gln Leu Gln Gly His Val Ala Pro
130 135 140

His Val Arg Ala Ile Ser Cys Leu Lys Gly Phe Glu Leu Gly Ser Lys
145 150 155 160

Gly Val Gln Leu Leu Ser Ser Tyr Val Thr Asp Glu Leu Gly Ile Gln
165 170 175

Cys Gly Ala Leu Ser Gly Ala Asn Leu Ala Pro Glu Val Ala Lys Glu
180 185 190

His Trp Ser Glu Thr Thr Val Ala Tyr Gln Leu Pro Lys Asp Tyr Gln
195 200 205

Gly Asp Gly Lys Asp Val Asp His Lys Ile Leu Lys Leu Leu Phe His
210 215 220

Arg Pro Tyr Phe His Val Asn Val Ile Asp Asp Val Ala Gly Ile Ser
225 230 235 240

Ile Ala Gly Ala Leu Lys Asn Val Val Ala Leu Ala Cys Gly Phe Val
245 250 255

Glu Gly Met Gly Trp Gly Asn Asn Ala Ser Ala Ala Ile Gln Arg Leu
260 265 270

Gly Leu Gly Glu Ile Ile Lys Phe Gly Arg Met Phe Phe Pro Glu Ser
275 280 285

Lys Val Glu Thr Tyr Tyr Gln Glu Ser Ala Gly Val Ala Asp Leu Ile
290 295 300

Thr Thr Cys Ser Gly Gly Arg Asn Val Lys Val Ala Thr Tyr Met Ala
305 310 315 320

Lys Thr Gly Lys Ser Ala Leu Glu Ala Glu Lys Glu Leu Leu Asn Gly
325 330 335

Gln Ser Ala Gln Gly Ile Ile Thr Cys Arg Glu Val His Glu Trp Leu
340 345 350

Gln Thr Cys Glu Leu Thr Gln Glu Phe Pro Ile Ile Arg Gly Ser Leu
355 360 365

Pro Asp Ser Leu Gln Gln Arg Pro His Gly Arg Pro Thr Gly Asp Asp
370 375 380

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 614 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: unknown
(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Thr Arg Ala Thr Trp Cys Asn Ser Pro Pro Pro Pro Leu His Arg Gln
1 5 10 15

Val Ser Arg Arg Asp Leu Leu Asp Arg Leu Asp Lys Thr His Gln Phe
20 25 30

Asp Val Leu Ile Ile Gly Gly Ala Thr Gly Thr Gly Cys Ala Leu
35 40 45

Asp Ala Ala Thr Arg Gly Leu Asn Val Ala Leu Val Glu Lys Gly Asp
50 55 60

Phe Ala Ser Gly Thr Ser Ser Lys Ser Thr Lys Met Ile His Gly Gly
65 70 75 80

Val Arg Tyr Leu Glu Lys Ala Phe Trp Glu Phe Ser Lys Ala Gln Leu
85 90 95

Asp Leu Val Ile Glu Ala Leu Asn Glu Arg Lys His Leu Ile Asn Thr
100 105 110

Ala Pro His Leu Cys Thr Val Leu Pro Ile Leu Ile Pro Ile Tyr Ser
115 120 125

Thr Trp Gln Val Pro Tyr Ile Tyr Met Gly Cys Lys Phe Tyr Asp Phe
130 135 140

Phe Gly Gly Ser Gln Asn Leu Lys Lys Ser Tyr Leu Leu Ser Lys Ser
145 150 155 160

Ala Thr Val Glu Lys Ala Pro Met Leu Thr Thr Asp Asn Leu Lys Ala
165 170 175

Ser Leu Val Tyr His Asp Gly Ser Phe Asn Asp Ser Arg Leu Asn Ala
180 185 190

Thr Leu Ala Ile Thr Gly Val Glu Asn Gly Ala Thr Val Leu Ile Tyr
195 200 205

Val Glu Val Gln Lys Leu Ile Lys Asp Pro Thr Ser Gly Lys Val Ile
210 215 220

Gly Ala Glu Ala Arg Asp Val Glu Thr Asn Glu Leu Val Arg Ile Asn
225 230 235 240

Ala Lys Cys Val Val Asn Ala Thr Gly Pro Tyr Ser Asp Ala Ile Leu
 245 250 255
 Gln Met Asp Arg Asn Pro Ser Gly Leu Pro Asp Ser Pro Leu Asn Asp
 260 265 270
 Asn Ser Lys Ile Lys Ser Thr Phe Asn Gln Ile Ser Val Met Asp Pro
 275 280 285
 Lys Met Val Ile Pro Ser Ile Gly Val His Ile Val Leu Pro Ser Phe
 290 295 300
 Tyr Ser Pro Lys Asp Met Gly Leu Leu Asp Val Arg Thr Ser Asp Gly
 305 310 315 320
 Arg Val Met Phe Phe Leu Pro Trp Gln Gly Lys Val Leu Ala Gly Thr
 325 330 335
 Thr Asp Ile Pro Leu Lys Gln Val Pro Glu Asn Pro Met Pro Thr Glu
 340 345 350
 Ala Asp Ile Gln Asp Ile Leu Lys Glu Leu Gln His Tyr Ile Glu Phe
 355 360 365
 Pro Val Lys Arg Glu Asp Val Leu Ser Ala Trp Ala Gly Val Arg Pro
 370 375 380
 Leu Val Arg Asp Pro Arg Thr Ile Pro Ala Asp Gly Lys Lys Gly Ser
 385 390 395 400
 Ala Thr Gln Gly Val Val Arg Ser His Phe Leu Phe Thr Ser Asp Asn
 405 410 415
 Gly Leu Ile Thr Ile Ala Gly Gly Lys Trp Thr Thr Tyr Arg Gln Met
 420 425 430
 Ala Glu Glu Thr Val Asp Lys Val Val Glu Val Gly Gly Phe His Asn
 435 440 445
 Leu Lys Pro Cys His Thr Arg Asp Ile Lys Leu Ala Gly Ala Glu Glu
 450 455 460
 Trp Thr Gln Asn Tyr Val Ala Leu Leu Ala Gln Asn Tyr His Leu Ser
 465 470 475 480
 Ser Lys Met Ser Asn Tyr Leu Val Gln Asn Tyr Gly Thr Arg Ser Ser
 485 490 495
 Ile Ile Cys Glu Phe Phe Lys Glu Ser Met Glu Asn Lys Leu Pro Leu
 500 505 510
 Ser Leu Ala Asp Lys Glu Asn Asn Val Ile Tyr Ser Ser Glu Glu Asn
 515 520 525
 Asn Leu Val Asn Phe Asp Thr Phe Arg Tyr Pro Phe Thr Ile Gly Glu
 530 535 540

Leu Lys Tyr Ser Met Gln Tyr Glu Tyr Cys Arg Thr Pro Leu Asp Phe
 545 550 555 560
 Leu Leu Arg Arg Thr Arg Phe Ala Phe Leu Asp Ala Lys Glu Ala Leu
 565 570 575
 Asn Ala Val His Ala Thr Val Lys Val Met Gly Asp Glu Phe Asn Trp
 580 585 590
 Ser Glu Lys Lys Arg Gln Trp Glu Leu Glu Lys Thr Val Asn Phe Ile
 595 600 605
 Gln Gly Arg Phe Gly Val
 610

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 339 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Asn Gln Arg Asn Ala Ser Met Thr Val Ile Gly Ala Gly Ser Tyr
 1 5 10 15
 Gly Thr Ala Leu Ala Ile Thr Leu Ala Arg Asn Gly His Glu Val Val
 20 25 30
 Leu Trp Gly His Asp Pro Glu His Ile Ala Thr Leu Glu Arg Asp Arg
 35 40 45
 Cys Asn Ala Ala Phe Leu Pro Asp Val Pro Phe Pro Asp Thr Leu His
 50 55 60
 Leu Glu Ser Asp Leu Ala Thr Ala Leu Ala Ala Ser Arg Asn Ile Leu
 65 70 75 80
 Val Val Val Pro Ser His Val Phe Gly Glu Val Leu Arg Gln Ile Lys
 85 90 95
 Pro Leu Met Arg Pro Asp Ala Arg Leu Val Trp Ala Thr Lys Gly Leu
 100 105 110
 Glu Ala Glu Thr Gly Arg Leu Leu Gln Asp Val Ala Arg Glu Ala Leu
 115 120 125
 Gly Asp Gln Ile Pro Leu Ala Val Ile Ser Gly Pro Thr Phe Ala Lys
 130 135 140
 Glu Leu Ala Ala Gly Leu Pro Thr Ala Ile Ser Leu Ala Ser Thr Asp
 145 150 155 160

Gln Thr Phe Ala Asp Asp Leu Gln Gln Leu Leu His Cys Gly Lys Ser
165 170 175

Phe Arg Val Tyr Ser Asn Pro Asp Phe Ile Gly Val Gln Leu Gly Gly
180 185 190

Ala Val Lys Asn Val Ile Ala Ile Gly Ala Gly Met Ser Asp Gly Ile
195 200 205

Gly Phe Gly Ala Asn Ala Arg Thr Ala Leu Ile Thr Arg Gly Leu Ala
210 215 220

Glu Met Ser Arg Leu Gly Ala Ala Leu Gly Ala Asp Pro Ala Thr Phe
225 230 235 240

Met Gly Met Ala Gly Leu Gly Asp Leu Val Leu Thr Cys Thr Asp Asn
245 250 255

Gln Ser Arg Asn Arg Arg Phe Gly Met Met Leu Gly Gln Gly Met Asp
260 265 270

Val Gln Ser Ala Gln Glu Lys Ile Gly Gln Val Val Glu Gly Tyr Arg
275 280 285

Asn Thr Lys Glu Val Arg Glu Leu Ala His Arg Phe Gly Val Glu Met
290 295 300

Pro Ile Thr Glu Glu Ile Tyr Gln Val Leu Tyr Cys Gly Lys Asn Ala
305 310 315 320

Arg Glu Ala Ala Leu Thr Leu Leu Gly Arg Ala Arg Lys Asp Glu Arg
325 330 335

Ser Ser His

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 501 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Met Glu Thr Lys Asp Leu Ile Val Ile Gly Gly Ile Asn Gly Ala
1 5 10 15

Gly Ile Ala Ala Asp Ala Ala Gly Arg Gly Leu Ser Val Leu Met Leu
20 25 30

Glu Ala Gln Asp Leu Ala Cys Ala Thr Ser Ser Ala Ser Ser Lys Leu
35 40 45

Ile His Gly Gly Leu Arg Tyr Leu Glu His Tyr Glu Phe Arg Leu Val
50 55 60

Ser Glu Ala Leu Ala Glu Arg Glu Val Leu Leu Lys Met Ala Pro His
 65 70 75 80
 Ile Ala Phe Pro Met Arg Phe Arg Leu Pro His Arg Pro His Leu Arg
 85 90 95
 Pro Ala Trp Met Ile Arg Ile Gly Leu Phe Met Tyr Asp His Leu Gly
 100 105 110
 Lys Arg Thr Ser Leu Pro Gly Ser Thr Gly Leu Arg Phe Gly Ala Asn
 115 120 125
 Ser Val Leu Lys Pro Glu Ile Lys Arg Gly Phe Glu Tyr Ser Asp Cys
 130 135 140
 Trp Val Asp Asp Ala Arg Leu Val Leu Ala Asn Ala Gln Met Val Val
 145 150 155 160
 Arg Lys Gly Gly Glu Val Leu Thr Arg Thr Arg Ala Thr Ser Ala Arg
 165 170 175
 Arg Glu Asn Gly Leu Trp Ile Val Glu Ala Glu Asp Ile Asp Thr Gly
 180 185 190
 Lys Lys Tyr Ser Trp Gln Ala Arg Gly Leu Val Asn Ala Thr Gly Pro
 195 200 205
 Trp Val Lys Gln Phe Phe Asp Asp Gly Met His Leu Pro Ser Pro Tyr
 210 215 220
 Gly Ile Arg Leu Ile Lys Gly Ser His Ile Val Val Pro Arg Val His
 225 230 235 240
 Thr Gln Lys Gln Ala Tyr Ile Leu Gln Asn Glu Asp Lys Arg Ile Val
 245 250 255
 Phe Val Ile Pro Trp Met Asp Glu Phe Ser Ile Ile Gly Thr Thr Asp
 260 265 270
 Val Glu Tyr Lys Gly Asp Pro Lys Ala Val Lys Ile Glu Glu Ser Glu
 275 280 285
 Ile Asn Tyr Leu Leu Asn Val Tyr Asn Thr His Phe Lys Lys Gln Leu
 290 295 300
 Ser Arg Asp Asp Ile Val Trp Thr Tyr Ser Gly Val Arg Pro Leu Cys
 305 310 315 320
 Asp Asp Glu Ser Asp Ser Pro Gln Ala Ile Thr Arg Asp Tyr Thr Leu
 325 330 335
 Asp Ile His Asp Glu Asn Gly Lys Ala Pro Leu Leu Ser Val Phe Gly
 340 345 350
 Gly Lys Leu Thr Thr Tyr Arg Lys Leu Ala Glu His Ala Leu Glu Lys
 355 360 365

Leu Thr Pro Tyr Tyr Gln Gly Ile Gly Pro Ala Trp Thr Lys Glu Ser
 370 375 380
 Val Leu Pro Gly Gly Ala Ile Glu Gly Asp Arg Asp Asp Tyr Ala Ala
 385 390 395 400
 Arg Leu Arg Arg Arg Tyr Pro Phe Leu Thr Glu Ser Leu Ala Arg His
 405 410 415
 Tyr Ala Arg Thr Tyr Gly Ser Asn Ser Glu Leu Leu Leu Gly Asn Ala
 420 425 430
 Gly Thr Val Ser Asp Leu Gly Glu Asp Phe Gly His Glu Phe Tyr Glu
 435 440 445
 Ala Glu Leu Lys Tyr Leu Val Asp His Glu Trp Val Arg Arg Ala Asp
 450 455 460
 Asp Ala Leu Trp Arg Arg Thr Lys Gln Gly Met Trp Leu Asn Ala Asp
 465 470 475 480
 Gln Gln Ser Arg Val Ser Gln Trp Leu Val Glu Tyr Thr Gln Gln Arg
 485 490 495
 Leu Ser Leu Ala Ser
 500

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 542 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Lys Thr Arg Asp Ser Gln Ser Ser Asp Val Ile Ile Ile Gly Gly
 1 5 10 15
 Gly Ala Thr Gly Ala Gly Ile Ala Arg Asp Cys Ala Leu Arg Gly Leu
 20 25 30
 Arg Val Ile Leu Val Glu Arg His Asp Ile Ala Thr Gly Ala Thr Gly
 35 40 45
 Arg Asn His Gly Leu Leu His Ser Gly Ala Arg Tyr Ala Val Thr Asp
 50 55 60
 Ala Glu Ser Ala Arg Glu Cys Ile Ser Glu Asn Gln Ile Leu Lys Arg
 65 70 75 80
 Ile Ala Arg His Cys Val Glu Pro Thr Asn Gly Leu Phe Ile Thr Leu
 85 90 95

Pro Glu Asp Asp Leu Ser Phe Gln Ala Thr Phe Ile Arg Ala Cys Glu
100 105 110

Glu Ala Gly Ile Ser Ala Glu Ala Ile Asp Pro Gln Gln Ala Arg Ile
115 120 125

Ile Glu Pro Ala Val Asn Pro Ala Leu Ile Gly Ala Val Lys Val Pro
130 135 140

Asp Gly Thr Val Asp Pro Phe Arg Leu Thr Ala Ala Asn Met Leu Asp
145 150 155 160

Ala Lys Glu His Gly Ala Val Ile Leu Thr Ala His Glu Val Thr Gly
165 170 175

Leu Ile Arg Glu Gly Ala Thr Val Cys Gly Val Arg Val Arg Asn His
180 185 190

Leu Thr Gly Glu Thr Gln Ala Leu His Ala Pro Val Val Val Asn Ala
195 200 205

Ala Gly Ile Trp Gly Gln His Ile Ala Glu Tyr Ala Asp Leu Arg Ile
210 215 220

Arg Met Phe Pro Ala Lys Gly Ser Leu Leu Ile Met Asp His Arg Ile
225 230 235 240

Asn Gln His Val Ile Asn Arg Cys Arg Lys Pro Ser Asp Ala Asp Ile
245 250 255

Leu Val Pro Gly Asp Thr Ile Ser Leu Ile Gly Thr Thr Ser Leu Arg
260 265 270

Ile Asp Tyr Asn Glu Ile Asp Asp Asn Arg Val Thr Ala Glu Glu Val
275 280 285

Asp Ile Leu Leu Arg Glu Gly Glu Lys Leu Ala Pro Val Met Ala Lys
290 295 300

Thr Arg Ile Leu Arg Ala Tyr Ser Gly Val Arg Pro Leu Val Ala Ser
305 310 315 320

Asp Asp Asp Pro Ser Gly Arg Asn Leu Ser Arg Gly Ile Val Leu Leu
325 330 335

Asp His Ala Glu Arg Asp Gly Leu Asp Gly Phe Ile Thr Ile Thr Gly
340 345 350

Gly Lys Leu Met Thr Tyr Arg Leu Met Ala Glu Trp Ala Thr Asp Ala
355 360 365

Val Cys Arg Lys Leu Gly Asn Thr Arg Pro Cys Thr Thr Ala Asp Leu
370 375 380

Ala Leu Pro Gly Ser Gln Glu Pro Ala Glu Val Thr Leu Arg Lys Val
385 390 395 400

Ile Ser Leu Pro Ala Pro Leu Arg Gly Ser Ala Val Tyr Arg His Gly
 405 410 415
 Asp Arg Thr Pro Ala Trp Leu Ser Glu Gly Arg Leu His Arg Ser Leu
 420 425 430
 Val Cys Glu Cys Glu Ala Val Thr Ala Gly Glu Val Gln Tyr Ala Val
 435 440 445
 Glu Asn Leu Asn Val Asn Ser Leu Leu Asp Leu Arg Arg Arg Thr Arg
 450 455 460
 Val Gly Met Gly Thr Cys Gln Gly Glu Leu Cys Ala Cys Arg Ala Ala
 465 470 475 480
 Gly Leu Leu Gln Arg Phe Asn Val Thr Thr Ser Ala Gln Ser Ile Glu
 485 490 495
 Gln Leu Ser Thr Phe Leu Asn Glu Arg Trp Lys Gly Val Gln Pro Ile
 500 505 510
 Ala Trp Gly Asp Ala Leu Arg Glu Ser Glu Phe Thr Arg Trp Val Tyr
 515 520 525
 Gln Gly Leu Cys Gly Leu Glu Lys Glu Gln Lys Asp Ala Leu
 530 535 540

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 250 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Met Gly Leu Thr Thr Lys Pro Leu Ser Leu Lys Val Asn Ala Ala Leu
 1 5 10 15
 Phe Asp Val Asp Gly Thr Ile Ile Ser Gln Pro Ala Ile Ala Ala
 20 25 30
 Phe Trp Arg Asp Phe Gly Lys Asp Lys Pro Tyr Phe Asp Ala Glu His
 35 40 45
 Val Ile Gln Val Ser His Gly Trp Arg Thr Phe Asp Ala Ile Ala Lys
 50 55 60
 Phe Ala Pro Asp Phe Ala Asn Glu Glu Tyr Val Asn Lys Leu Glu Ala
 65 70 75 80
 Glu Ile Pro Val Lys Tyr Gly Glu Lys Ser Ile Glu Val Pro Gly Ala
 85 90 95

Val Lys Leu Cys Asn Ala Leu Asn Ala Leu Pro Lys Glu Lys Trp Ala
 100 105 110
 Val Ala Thr Ser Gly Thr Arg Asp Met Ala Gln Lys Trp Phe Glu His
 115 120 125
 Leu Gly Ile Arg Arg Pro Lys Tyr Phe Ile Thr Ala Asn Asp Val Lys
 130 135 140
 Gln Gly Lys Pro His Pro Glu Pro Tyr Leu Lys Gly Arg Asn Gly Leu
 145 150 155 160
 Gly Tyr Pro Ile Asn Glu Gln Asp Pro Ser Lys Ser Lys Val Val Val
 165 170 175
 Phe Glu Asp Ala Pro Ala Gly Ile Ala Ala Gly Lys Ala Ala Gly Cys
 180 185 190
 Lys Ile Ile Gly Ile Ala Thr Thr Phe Asp Leu Asp Phe Leu Lys Glu
 195 200 205
 Lys Gly Cys Asp Ile Ile Val Lys Asn His Glu Ser Ile Arg Val Gly
 210 215 220
 Gly Tyr Asn Ala Glu Thr Asp Glu Val Glu Phe Ile Phe Asp Asp Tyr
 225 230 235 240
 Leu Tyr Ala Lys Asp Asp Leu Leu Lys Trp
 245 250

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 271 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Lys Arg Phe Asn Val Leu Lys Tyr Ile Arg Thr Thr Lys Ala Asn
 1 5 10 15
 Ile Gln Thr Ile Ala Met Pro Leu Thr Thr Lys Pro Leu Ser Leu Lys
 20 25 30
 Ile Asn Ala Ala Leu Phe Asp Val Asp Gly Thr Ile Ile Ser Gln
 35 40 45
 Pro Ala Ile Ala Ala Phe Trp Arg Asp Phe Gly Lys Asp Lys Pro Tyr
 50 55 60
 Phe Asp Ala Glu His Val Ile His Ile Ser His Gly Trp Arg Thr Tyr
 65 70 75 80

Asp Ala Ile Ala Lys Phe Ala Pro Asp Phe Ala Asp Glu Glu Tyr Val
 85 90 95
 Asn Lys Leu Glu Gly Glu Ile Pro Glu Lys Tyr Gly Glu His Ser Ile
 100 105 110
 Glu Val Pro Gly Ala Val Lys Leu Cys Asn Ala Leu Asn Ala Leu Pro
 115 120 125
 Lys Glu Lys Trp Ala Val Ala Thr Ser Gly Thr Arg Asp Met Ala Lys
 130 135 140
 Lys Trp Phe Asp Ile Leu Lys Ile Lys Arg Pro Glu Tyr Phe Ile Thr
 145 150 155 160
 Ala Asn Asp Val Lys Gln Gly Lys Pro His Pro Glu Pro Tyr Leu Lys
 165 170 175
 Gly Arg Asn Gly Leu Gly Phe Pro Ile Asn Glu Gln Asp Pro Ser Lys
 180 185 190
 Ser Lys Val Val Val Phe Glu Asp Ala Pro Ala Gly Ile Ala Ala Gly
 195 200 205
 Lys Ala Ala Gly Cys Lys Ile Val Gly Ile Ala Thr Thr Phe Asp Leu
 210 215 220
 Asp Phe Leu Lys Glu Lys Gly Cys Asp Ile Ile Val Lys Asn His Glu
 225 230 235 240
 Ser Ile Arg Val Gly Glu Tyr Asn Ala Glu Thr Asp Glu Val Glu Leu
 245 250 255
 Ile Phe Asp Asp Tyr Leu Tyr Ala Lys Asp Asp Leu Leu Lys Trp
 260 265 270

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 709 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Met Phe Pro Ser Leu Phe Arg Leu Val Val Phe Ser Lys Arg Tyr Ile
 1 5 10 15
 Phe Arg Ser Ser Gln Arg Leu Tyr Thr Ser Leu Lys Gln Glu Gln Ser
 20 25 30
 Arg Met Ser Lys Ile Met Glu Asp Leu Arg Ser Asp Tyr Val Pro Leu
 35 40 45

Ile Ala Ser Ile Asp Val Gly Thr Thr Ser Ser Arg Cys Ile Leu Phe
 50 55 60
 Asn Arg Trp Gly Gln Asp Val Ser Lys His Gln Ile Glu Tyr Ser Thr
 65 70 75 80
 Ser Ala Ser Lys Gly Lys Ile Gly Val Ser Gly Leu Arg Arg Pro Ser
 85 90 95
 Thr Ala Pro Ala Arg Glu Thr Pro Asn Ala Gly Asp Ile Lys Thr Ser
 100 105 110
 Gly Lys Pro Ile Phe Ser Ala Glu Gly Tyr Ala Ile Gln Glu Thr Lys
 115 120 125
 Phe Leu Lys Ile Glu Glu Leu Asp Leu Asp Phe His Asn Glu Pro Thr
 130 135 140
 Leu Lys Phe Pro Lys Pro Gly Trp Val Glu Cys His Pro Gln Lys Leu
 145 150 155 160
 Leu Val Asn Val Val Gln Cys Leu Ala Ser Ser Leu Leu Ser Leu Gln
 165 170 175
 Thr Ile Asn Ser Glu Arg Val Ala Asn Gly Leu Pro Pro Tyr Lys Val
 180 185 190
 Ile Cys Met Gly Ile Ala Asn Met Arg Glu Thr Thr Ile Leu Trp Ser
 195 200 205
 Arg Arg Thr Gly Lys Pro Ile Val Asn Tyr Gly Ile Val Trp Asn Asp
 210 215 220
 Thr Arg Thr Ile Lys Ile Val Arg Asp Lys Trp Gln Asn Thr Ser Val
 225 230 235 240
 Asp Arg Gln Leu Gln Leu Arg Gln Lys Thr Gly Leu Pro Leu Leu Ser
 245 250 255
 Thr Tyr Phe Ser Cys Ser Lys Leu Arg Trp Phe Leu Asp Asn Glu Pro
 260 265 270
 Leu Cys Thr Lys Ala Tyr Glu Asn Asp Leu Met Phe Gly Thr Val
 275 280 285
 Asp Thr Trp Leu Ile Tyr Gln Leu Thr Lys Gln Lys Ala Phe Val Ser
 290 295 300
 Asp Val Thr Asn Ala Ser Arg Thr Gly Phe Met Asn Leu Ser Thr Leu
 305 310 315 320
 Lys Tyr Asp Asn Glu Leu Leu Glu Phe Trp Gly Ile Asp Lys Asn Leu
 325 330 335
 Ile His Met Pro Glu Ile Val Ser Ser Gln Tyr Tyr Gly Asp Phe
 340 345 350

Val	Leu	Arg	Asp	Leu	Val	Lys	Arg	Asn	Leu	Pro	Ile	Gln	Gly	Cys	Leu
370						375					380				
Gly	Asp	Gln	Ser	Ala	Ser	Met	Val	Gly	Gln	Leu	Ala	Tyr	Lys	Pro	Gly
385						390					395				400
Ala	Ala	Lys	Cys	Thr	Tyr	Gly	Thr	Gly	Cys	Phe	Leu	Leu	Tyr	Asn	Thr
											410				415
Gly	Thr	Lys	Lys	Leu	Ile	Ser	Gln	His	Gly	Ala	Leu	Thr	Thr	Leu	Ala
						420					425				430
Phe	Trp	Phe	Pro	His	Leu	Gln	Glu	Tyr	Gly	Gly	Gln	Lys	Pro	Glu	Leu
						435					440				445
Ser	Lys	Pro	His	Phe	Ala	Leu	Glu	Gly	Ser	Val	Ala	Val	Ala	Gly	Ala
						450					455				460
Val	Val	Gln	Trp	Leu	Arg	Asp	Asn	Leu	Arg	Leu	Ile	Asp	Lys	Ser	Glu
						465					470				480
Asp	Val	Gly	Pro	Ile	Ala	Ser	Thr	Val	Pro	Asp	Ser	Gly	Gly	Val	Val
						485					490				495
Phe	Val	Pro	Ala	Phe	Ser	Gly	Leu	Phe	Ala	Pro	Tyr	Trp	Asp	Pro	Asp
						500					505				510
Ala	Arg	Ala	Thr	Ile	Met	Gly	Met	Ser	Gln	Phe	Thr	Thr	Ala	Ser	His
						515					520				525
Ile	Ala	Arg	Ala	Ala	Ala	Val	Glu	Gly	Val	Cys	Phe	Gln	Ala	Arg	Ala
						530					535				540
Leu	Lys	Ala	Met	Ser	Ser	Asp	Ala	Phe	Gly	Glu	Gly	Ser	Lys	Asp	Arg
						545					550				560
Asp	Phe	Leu	Glu	Glu	Ile	Ser	Asp	Val	Thr	Tyr	Glu	Lys	Ser	Pro	Leu
						565					570				575
Ser	Val	Leu	Ala	Val	Asp	Gly	Gly	Met	Ser	Arg	Ser	Asn	Glu	Val	Met
						580					585				590
Gln	Ile	Gln	Ala	Asp	Ile	Leu	Gly	Pro	Cys	Val	Lys	Val	Arg	Arg	Ser
						595					600				605
Pro	Thr	Ala	Glu	Cys	Thr	Ala	Leu	Gly	Ala	Ala	Ile	Ala	Ala	Asn	Met
						610					615				620
Ala	Phe	Lys	Asp	Val	Asn	Glu	Arg	Pro	Leu	Trp	Lys	Asp	Leu	His	Asp
						625					630				640
Val	Lys	Lys	Trp	Val	Phe	Tyr	Asn	Gly	Met	Glu	Lys	Asn	Glu	Gln	Ile
						645					650				655

Ser Pro Glu Ala His Pro Asn Leu Lys Ile Phe Arg Ser Glu Ser Asp
660 665 670

Asp Ala Glu Arg Arg Lys His Trp Lys Tyr Trp Glu Val Ala Val Glu
675 680 685

Arg Ser Lys Gly Trp Leu Lys Asp Ile Glu Gly Glu His Glu Gln Val
690 695 700

Leu Glu Asn Phe Gln
705

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 51 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

GCGCGGATCC AGGAGTCTAG AATTATGGGA TTGACTACTA AACCTCTATC T 51

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 36 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

GATACGCCCG GGT TACCA TT TCAACAGATC GTC CTT 36

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

TTGATAATAT AACCATGGCT GCTGCTGCTG ATAG

34

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 39 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

GTATGATATG TTATCTTGGAA TCCAATAAAAT CTAATCTTC

39

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

CATGACTAGT AAGGAGGACAA ATTCA

24

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

CATGGAATTG TCCTCCCTTAC TAGT

24

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

CTAGTAAGGA GGACAATTCA

19

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

CATGGAATTG TCCTCCTTA

19

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

GATCCAGGAA ACAGA

15

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

CTAGTCTGTT TCCTG

15

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 22 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

GCTTTCTGTG CTGCGGCTTT AG

22

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

TGGTCGAGGA TCCACTTCAC TTT

23

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 51 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

AAAGTGAAGT GGATCCTCGA CCAATTGGAT GGTGGCGCAG TAGCAAACAA T

51

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

GGATCACCGC CGCAGAAACT ACG

23

(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

CTGTCAGCCG TTAAGTGTTC CTGTG

25

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

CAGTTCAACC TGTTGATAAGT ACG

23

(2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

ATGAGTCAAA CATCAACCTT

20

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

ATGGAGAAAA AAATCACTGG

20

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

TTACGCCCG CCCTGCCACT

20

(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "PRIMER"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

TCAGAGGATG TGCACCTGCA

20

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "PRIMER"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

CGAGCATGCC GCATTTGGCA CTACTC

26

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "PRIMER"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

GCGTCTAGAG TAGGTTATTC CCACTCTG

29

(2) INFORMATION FOR SEQ ID NO:38:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 26 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "PRIMER"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

GAAGTCGACC GCTGCGCCTT ATCCGG

26

(2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 base pairs

(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

CGCGTCGACG TTTACAATTT CAGGTGGC

28

(2) INFORMATION FOR SEQ ID NO:40:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

GCAGCATGCT GGAATGGTAG TAG

23

(2) INFORMATION FOR SEQ ID NO:41:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

CAGTCTAGAG TTATTGGCAA ACCTACC

27

(2) INFORMATION FOR SEQ ID NO:42:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

GATGCATGCC CAGGGCGGAG ACGGC

25

(2) INFORMATION FOR SEQ ID NO:43:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 29 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PRIMER"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

CTAACGATTG TTCTCTAGAG AAAATGTCC

29

Sequence Readings
Sequence Readings
Sequence Readings
Sequence Readings
Sequence Readings

WHAT IS CLAIMED IS:

1. A method for the production of glycerol from a recombinant organism comprising:
 - (i) transforming a suitable host cell with an expression cassette comprising either one or both of
 - 5 (a) a gene encoding a protein having glycerol-3-phosphate dehydrogenase activity, and
 - (b) a gene encoding a protein having glycerol-3-phosphate phosphatase activity,
- 10 the suitable host cell having a disruption in either one or both of
 - (a) an endogenous gene encoding a polypeptide having glycerol kinase activity, and
 - (b) an endogenous gene encoding a polypeptide having glycerol dehydrogenase activity,
- 15 wherein the disruption prevents the expression of active gene product;
 - (ii) culturing the transformed host cell of (i) in the presence of at least one carbon source selected from the group consisting of monosaccharides, oligosaccharides, polysaccharides, and single-carbon substrates, whereby glycerol is produced; and
 - 20 (iii) optionally recovering the glycerol produced in (ii).
2. The method of Claim 1 wherein the expression cassette comprises a gene encoding a glycerol-3-phosphate dehydrogenase enzyme.
3. The method of Claim 1 wherein the expression cassette comprises a gene encoding a glycerol-3-phosphate phosphatase enzyme.
- 25 4. The method of Claim 1 wherein the expression cassette comprises genes encoding a glycerol-3-phosphate phosphatase enzyme and a glycerol-3-phosphate dehydrogenase enzyme.
5. The method of Claim 1 wherein the host cell contains a disruption in a gene encoding an endogenous glycerol kinase enzyme wherein the disruption prevents the expression of active gene product.
- 30 6. The method of Claim 1 wherein the host cell contains a disruption in a gene encoding an endogenous glycerol dehydrogenase enzyme wherein the disruption prevents the expression of active gene product.
7. The method of Claim 1 wherein the host cell contains a) a disruption in a gene encoding an endogenous glycerol kinase enzyme and b) a disruption in a gene encoding an endogenous glycerol dehydrogenase enzyme, wherein the disruptions in the respective genes prevent the expression of active gene product from either gene.

8. The method of Claim 1 wherein the suitable host cell is selected from the group consisting of bacteria, yeast, and filamentous fungi.

9. The method of Claim 8 wherein the suitable host cell is selected from the group consisting of *Citrobacter*, *Enterobacter*, *Clostridium*, *Klebsiella*,
5 *Aerobacter*, *Lactobacillus*, *Aspergillus*, *Saccharomyces*, *Schizosaccharomyces*,
Zygosaccharomyces, *Pichia*, *Kluyveromyces*, *Candida*, *Hansenula*,
Debaryomyces, *Mucor*, *Torulopsis*, *Methylobacter*, *Escherichia*, *Salmonella*,
Bacillus, *Streptomyces*, and *Pseudomonas*.

10. The method of Claim 9 wherein the suitable host cell is *E. coli* or
10 *Saccharomyces sp.*

11. The method of Claim 1 wherein the carbon source is glucose.

12. The method of Claim 1 wherein the protein having glycerol-3-phosphate dehydrogenase activity corresponds to amino acid sequences selected from the group consisting of SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9,
15 SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12 and wherein the amino acid sequences encompasses amino acid substitutions, deletions or insertions that do not alter the functional properties of the enzyme.

13. The method of Claim 1 wherein the protein having glycerol-3-phosphatase activity corresponds to the amino acid sequences selected from the group consisting of SEQ ID NO:13 and SEQ ID NO:14, and wherein the amino acid sequences may encompass amino acid substitutions, deletions or additions that do not alter the function of the enzyme.

14. A transformed host cell comprising:

25 (a) a gene encoding a protein having a glycerol-3-phosphate dehydrogenase activity;

(b) a gene encoding a protein having glycerol-3-phosphate phosphatase activity;

(c) a disruption in a gene encoding an endogenous glycerol kinase and;

30 (d) a disruption a gene encoding an endogenous glycerol dehydrogenase;

wherein the disruptions in the genes of (c) and (d) prevent the expression of active gene product, and wherein the host cell converts at least one carbon source selected from the group consisting of monosaccharides, oligosaccharides, polysaccharides, and single-carbon substrates to glycerol.

35 15. A transformed host cell comprising:

(a) a gene encoding a protein having a glycerol-3-phosphate dehydrogenase activity;

(b) a gene encoding a protein having glycerol-3-phosphate phosphatase activity; and

(c) a disruption in a gene encoding an endogenous glycerol dehydrogenase;

5 wherein the disruption in the gene of (c) prevents the expression of active gene product, and wherein the host cell converts at least one carbon source selected from the group consisting of monosaccharides, oligosaccharides, polysaccharides, and single-carbon substrates to glycerol.

16. A transformed host cell comprising:

10 (a) a gene encoding a protein having a glycerol-3-phosphate dehydrogenase activity;

 (b) a gene encoding a protein having glycerol-3-phosphate phosphatase activity; and

 (c) a disruption in a gene encoding an endogenous glycerol

15 kinase,

 wherein the disruption in the gene of (c) prevents the expression of active gene product, and wherein the host cell converts at least one carbon source selected from the group consisting of monosaccharides, oligosaccharides, polysaccharides, and single-carbon substrates to glycerol.

20 17. A method for the production of 1,3-propanediol from a recombinant organism comprising:

 (i) transforming a suitable host cell with an expression cassette comprising either one or both of

 (a) a gene encoding a protein having glycerol-3-phosphate dehydrogenase activity, and

 (b) a gene encoding a protein having glycerol-3-phosphate phosphatase activity,

 the suitable host cell having at least one gene encoding a protein having a dehydratase activity and having a disruption in either one or both of:

30 (a) an endogenous gene encoding a polypeptide having glycerol kinase activity, and

 (b) an endogenous gene encoding a polypeptide having glycerol dehydrogenase activity,

 wherein the disruption in the genes of (a) or (b) prevents the expression of active gene product;

35 (ii) culturing the transformed host cell of (i) in the presence of at least one carbon source selected from the group consisting of monosaccharides, oligosaccharides, polysaccharides, and single-carbon substrates whereby 1,3-propanediol is produced; and

(iii) recovering the 1,3-propanediol produced in (ii).

18. The method of Claim 17 wherein the protein having a dehydratase activity is selected from the group consisting of a glycerol dehydratase enzyme and a diol dehydratase enzyme.

5 19. The method of Claim 18 wherein the glycerol dehydratase enzyme is encoded by a gene, the gene isolated from a microorganism, the microorganism selected from the group consisting of *Klebsiella*, *Lactobacillus*, *Enterobacter*, *Citrobacter*, *Pelobacter*, *Ityobacter*, and *Clostridium*.

20. The method of Claim 18 wherein the diol dehydratase enzyme is
10 encoded by a gene, the gene isolated from a microorganism, the microorganism selected from the group consisting of *Klebsiella* and *Salmonella*.

TITLE

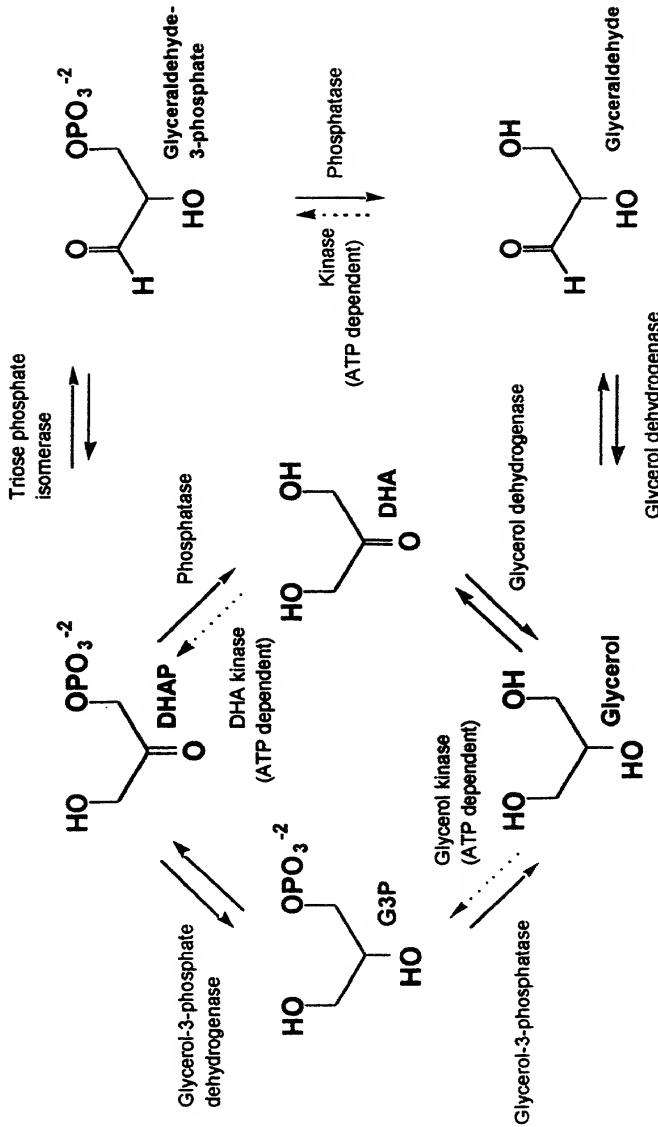
METHOD FOR THE PRODUCTION OF GLYCEROL
BY RECOMBINANT ORGANISMS

ABSTRACT OF INVENTION

5 Recombinant organisms are provided comprising genes encoding a glycerol-3-phosphate dehydrogenase and/or a glycerol-3-phosphatase activity useful for the production of glycerol from a variety of carbon substrates. The organisms further contain disruptions in the endogenous genes encoding proteins having glycerol kinase and glycerol dehydrogenase activities.

10

FIG. 1



PATENT
EXPRESS MAIL LABEL NO. EL073739163US

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In the Application of:

RAMESH V. NAIR ET AL. CASE NO.: CR9981 US DIV

APPLICATION NO.: UNKNOWN GROUP ART UNIT: UNKNOWN

FILED: CONCURRENTLY HEREWITH EXAMINER: UNKNOWN

FOR: METHOD FOR THE PRODUCTION OF
GLYCEROL BY RECOMBINANT ORGANISMS

DECLARATION IN ACCORDANCE WITH 37 CFR 1.821

Assistant Commissioner for Patents
Washington, D.C. 20231

Sir:

I hereby state that the content of the paper and computer
readable copies of the Sequence Listing, submitted in
accordance with 37 CFR 1.821(c) and (e), respectively are the
same.

Respectfully submitted,

Linda Axamethy Floyd
LINDA AXAMETHY FLOYD
ATTORNEY FOR APPLICANTS
REGISTRATION NO. 33,692
TELEPHONE: 302-892-8112

Dated: 25 October 2000

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

IN THE APPLICATION OF:

RAMESH V. NAIR ET AL.

APPLICATION NO: UNKNOWN

FILED: CONCURRENTLY HEREWITH

CASE NO.: CR9981 US DIV

TITLE: METHOD FOR THE PRODUCTION OF GLYCEROL BY
RECOMBINANT ORGANISMSAssistant Commissioner for Patents
Washington, D.C. 20231**DECLARATION OF BIOLOGICAL CULTURE DEPOSIT**

Sir:

I, Linda Axamethy Floyd, declare that:

I am an attorney of record for the owner of the above-identified application.

Cultures of the following biological materials have been deposited with the following international depository:

American Type Culture Collection (ATCC)
 10801 University Blvd.
 Manassas, VA 20110-2209, U.S.A.

under conditions that satisfy the requirements of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. A copy of the receipt issued pursuant to Rules 7.3 and 10.2 (stating the term of the deposit) is attached, except for ATCC 209120, the Budapest Treaty receipt for which will be provided at a later date.

Depositor's Identification of Organism	International Depository Accession Number	Date of Deposit
<i>Escherichia coli</i> RJF 10M	ATCC 98597	November 25, 1997
<i>E. coli</i> MSP 33.6	ATCC 98598	November 25, 1997
<i>Escherichia coli</i> DH5 α pAH21	ATCC 98187	September 26, 1996
<i>Escherichia coli</i> pDARIA/AA200	ATCC 98248	November 6, 1996

I further aver that all restrictions on the availability to the public of the culture will be irrevocably removed upon the granting of a U.S. patent on the above-identified application.

Respectfully submitted,

Linda Axamethy Floyd
 LINDA AXAMETHY FLOYD
 ATTORNEY FOR APPLICANTS
 US PTO REGISTRATION NO. 33,692
 TELEPHONE: (302) 892-8112

Dated:

Enclosure: Deposit Receipt

DECLARATION and POWER OF ATTORNEY

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

METHOD FOR THE PRODUCTION OF GLYCEROL BY RECOMBINANT ORGANISMS

the specification of which is attached hereto unless the following box is checked:

was filed on December 2, 1993, Application No. 08/982,783 or PCT International Application No. _____ and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is known to me to be material to patentability as defined in 37 CFR § 1.56.

I hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed.

Application No.	Country	Filing Date	Priority Claimed (Yes/No)
-----------------	---------	-------------	---------------------------

I hereby claim the benefit under 35 U.S.C. § 119(e) of any United States Provisional Application(s) listed below.

U.S. Provisional Application No.	U.S. Filing Date
----------------------------------	------------------

I hereby claim the benefit under 35 U.S.C. § 120 of any United States application(s), or § 365(c) of any PCT International Application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application or PCT International Application in the manner provided by the first paragraph of 35 U.S.C. § 112, I acknowledge the duty to disclose information which is known to me to be material to patentability as defined in 37 CFR § 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application.

Application No.	Filing Date	Status (patented, pending or abandoned)
-----------------	-------------	---

POWER OF ATTORNEY: I hereby appoint the following attorney(s) and/or agent(s) the power to prosecute this application and transact all business in the Patent and Trademark Office connected therewith:

Name: LINDA AXAMETHY FLOYD BARBARA SIEGELL	Registration No.: 33,692 30,684	
Send correspondence and direct telephone calls to: LINDA AXAMETHY FLOYD	E. I. du Pont de Nemours and Company Legal - Patents Wilmington, DE 19898, U.S.A.	Tel. No. (302) 892-8112

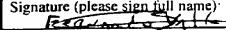
I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

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DECLARATION AND POWER OF ATTORNEY - Page 2

Docket No.: CR-9981-B

Full Name of Inventor	Last Name VALLE	First Name FERNANDO	Middle Name
	Signature (please sign full name): 		Date: 3-13-98
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Post Office Address	Post Office Address 1015 CADILLAC WAY, APT 212	City BURLINGAME	State or Country CALIFORNIA
			Zip Code 94010

SCANNED, #